



UNIVERSITY OF
CAMBRIDGE

Protein Structure Prediction and Using AlphaFold

Day 10: Friday 31st March

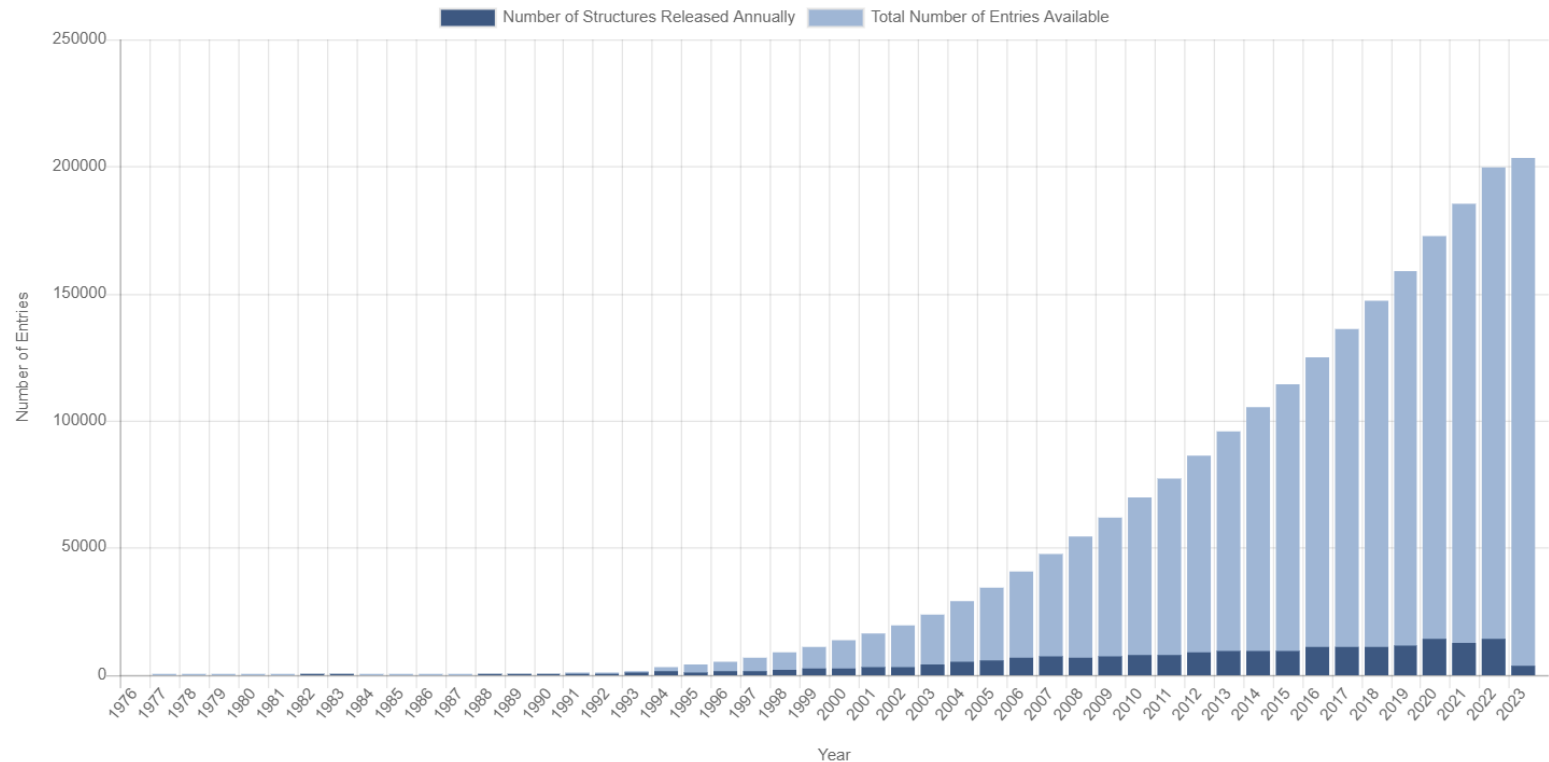
3D Structure Prediction

- Yesterday we learnt about experimental approaches to determine protein structure:
 - NMR
 - X-ray crystallography
 - Electron microscopy (cryo-EM)
- Today we learn about in silico approaches to predict structures:
 - Homology Modelling
 - Artificial Intelligence and AlphaFold
 - What AlphaFold can and can't do (yet)



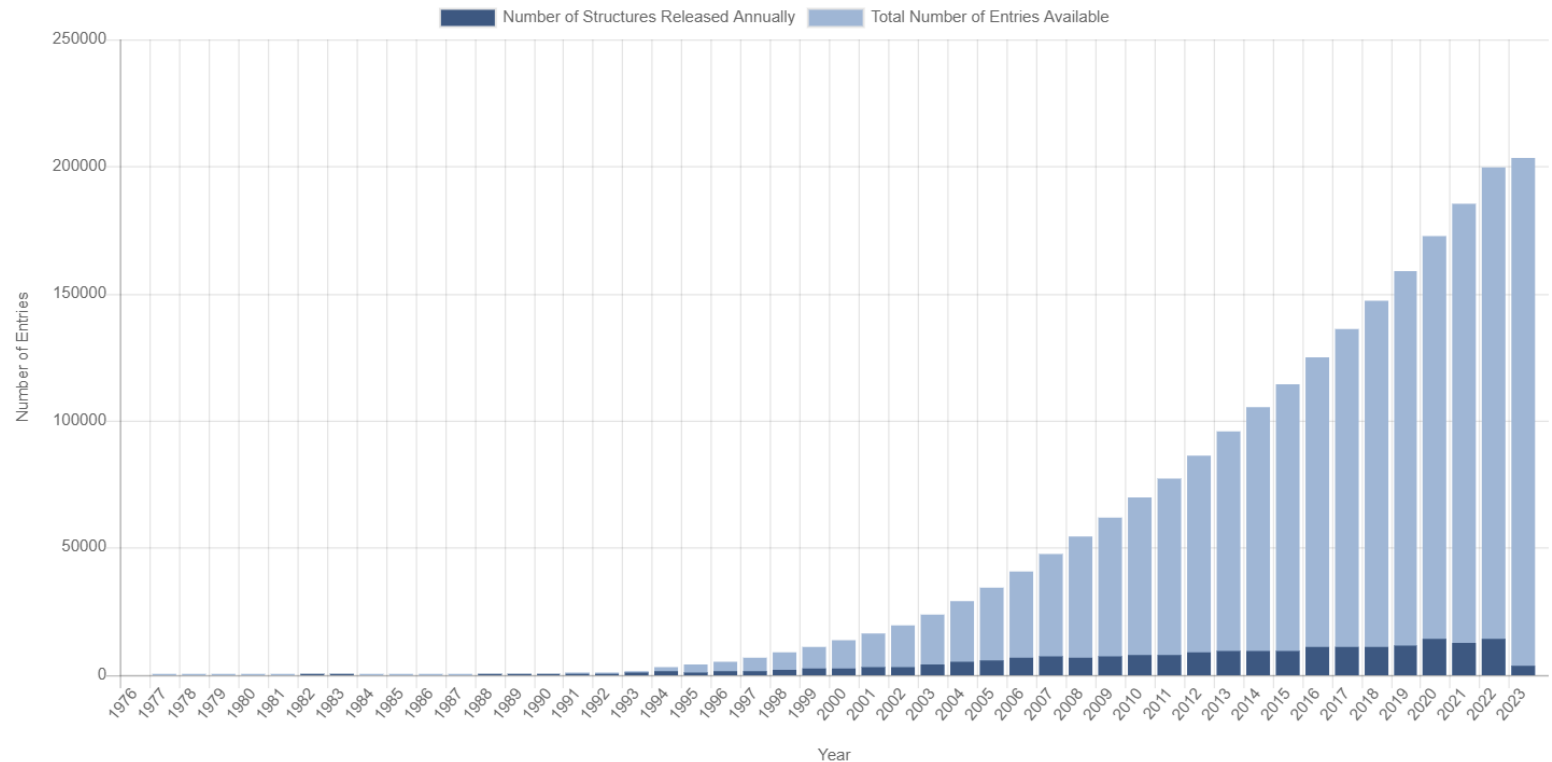
Experimental Structures in the PDB

- Enormous and growing number of structures that have been experimentally determined
- These are freely available in the online Protein Data Bank and listed in UniProt
- If an experimental structure exists – USE IT!



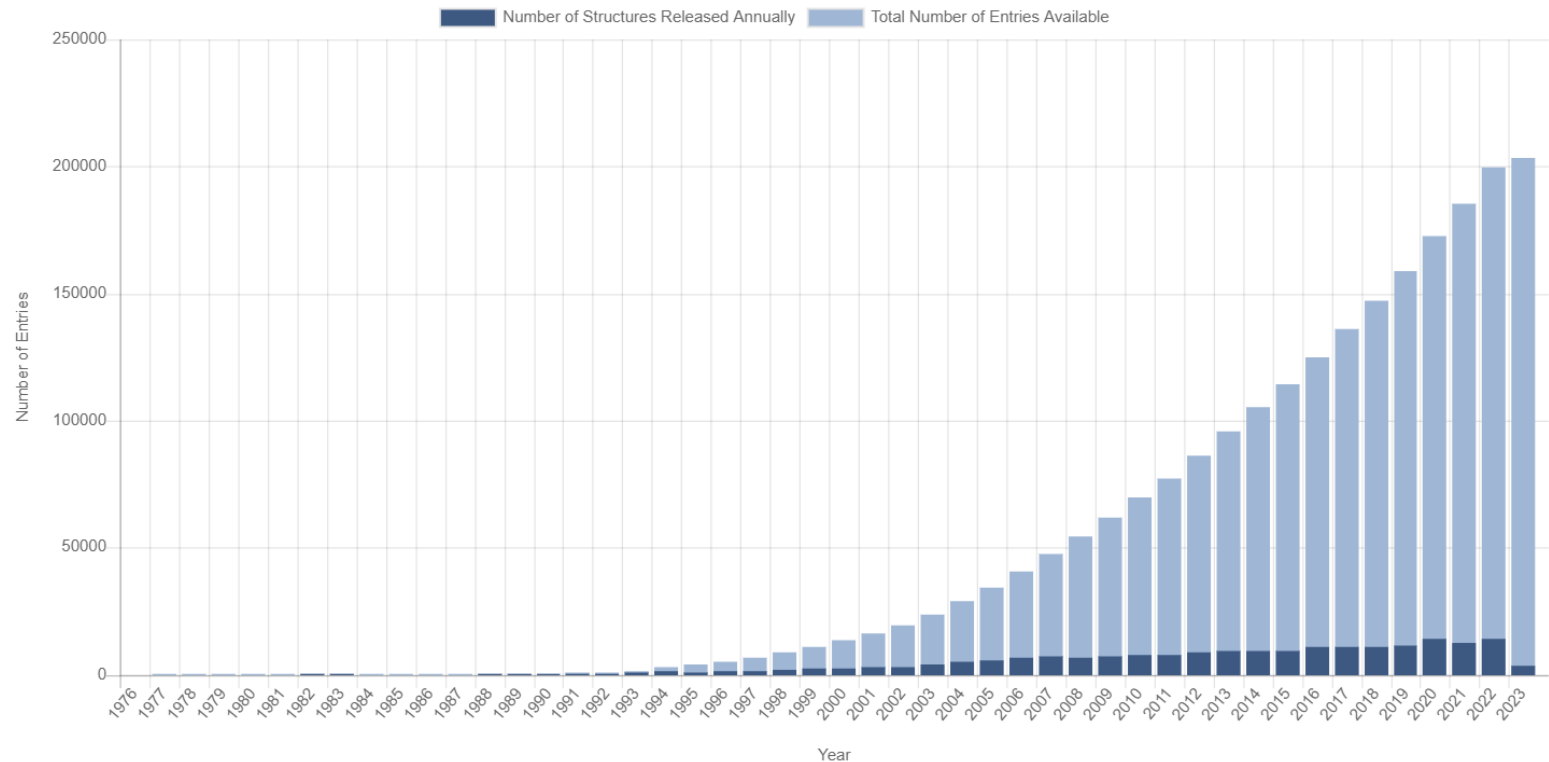
Experimental Structures in the PDB

- These experimental structures have been a very rich source of information for structure prediction for decades



Experimental Structures in the PDB

- These experimental structures have been a very rich source of information for structure prediction for decades
- This approach is called Homology Modelling



Predicting a 3D structure

- Homology modelling has existed for a long time – use a closely related known structure to predict a new one
 - Modeller
 - SwissModel
 - HHPred
 - FFAS
 - SCWRL



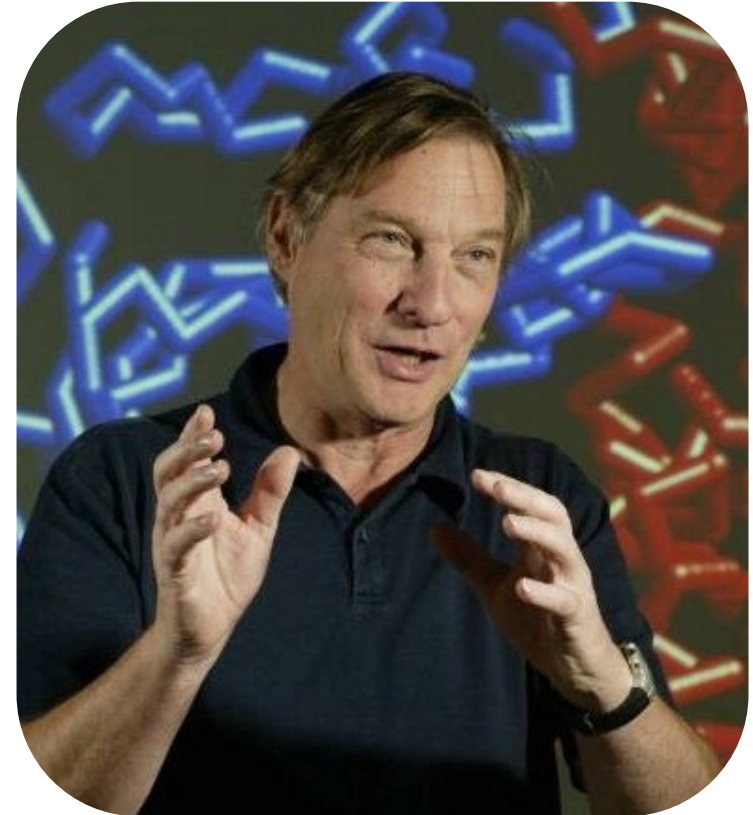
Predicting a 3D structure

- Homology modelling has existed for a long time – use a closely related known structure to predict a new one
 - Modeller
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 - SCWRL
- Ab initio modelling has been a huge challenge and actually stimulated an open competition called CASP

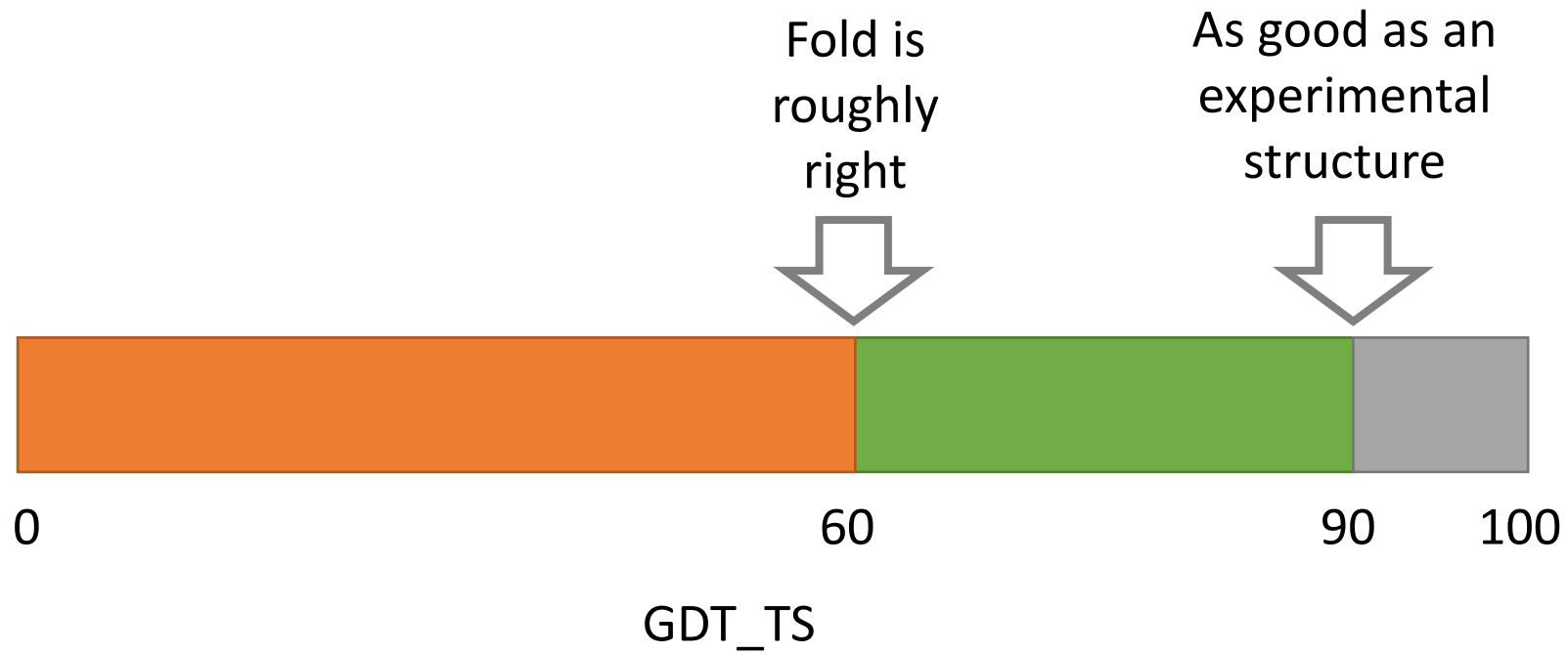


Critical Assessment of protein Structure Prediction (CASP)

- Experiment initiated by John Moult started in 1994
- Independent benchmark of ability to predict novel protein structures
- Targets ranked based on 'difficulty'
- Scored on accuracy and coverage of backbone prediction (GDT_TS)

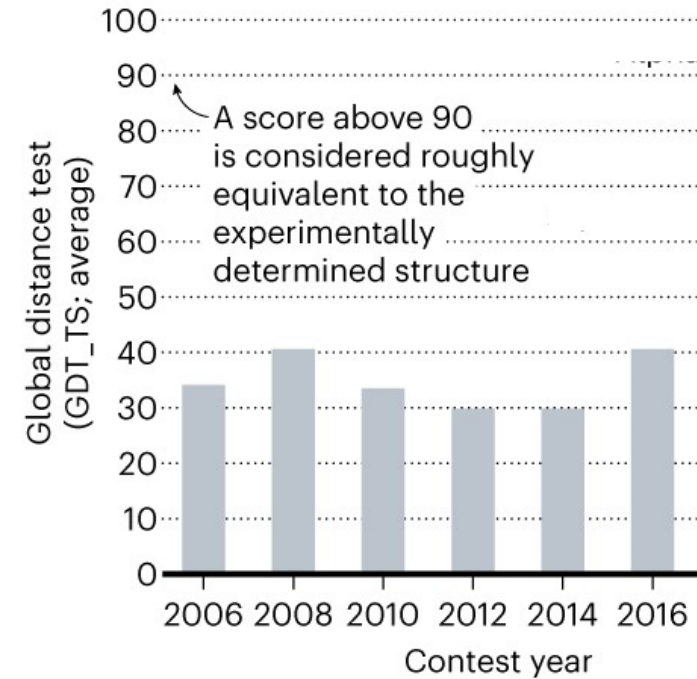
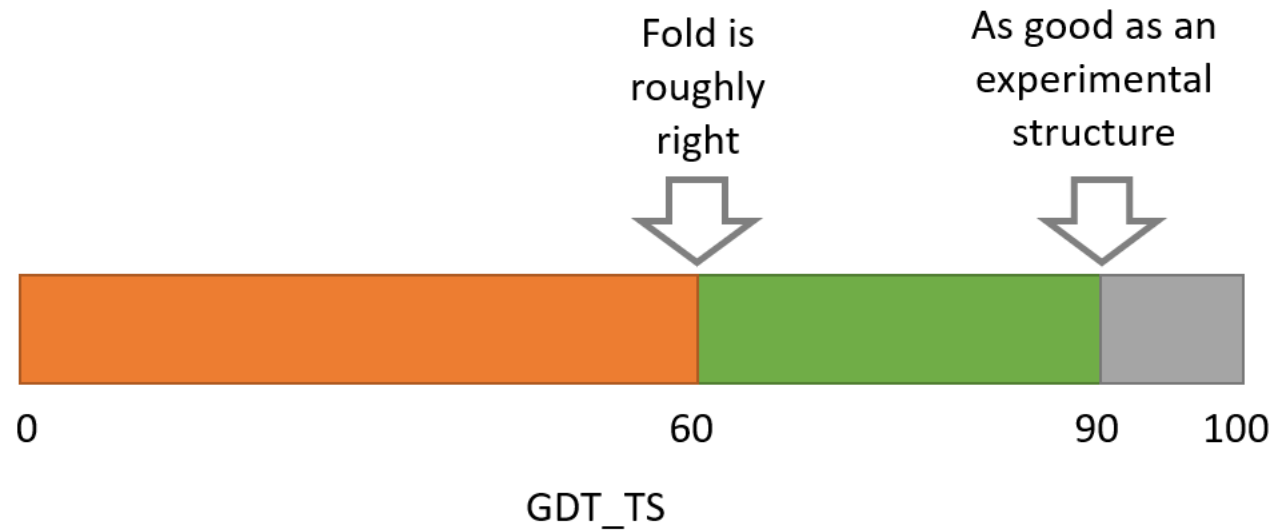


How good is the prediction?



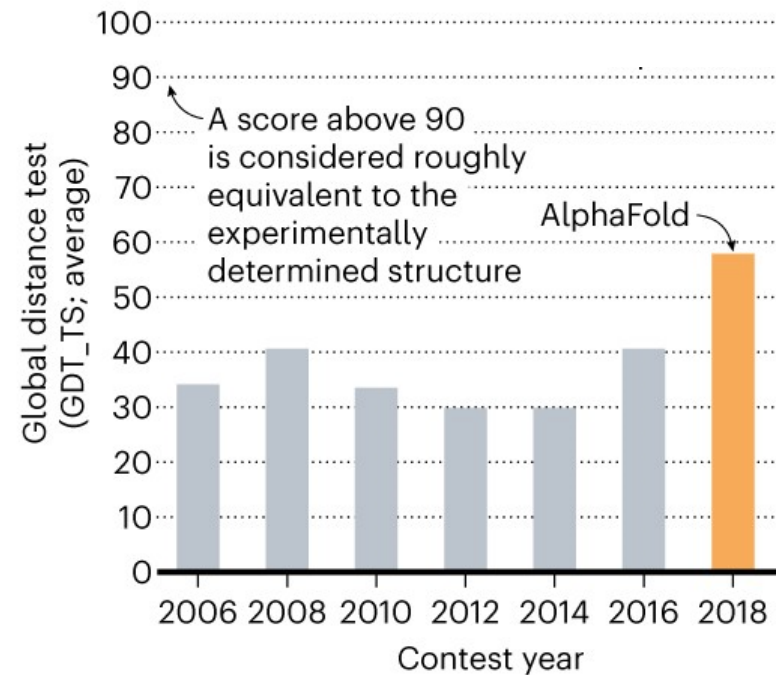
Predicting structures

- CASP outcomes and scoring



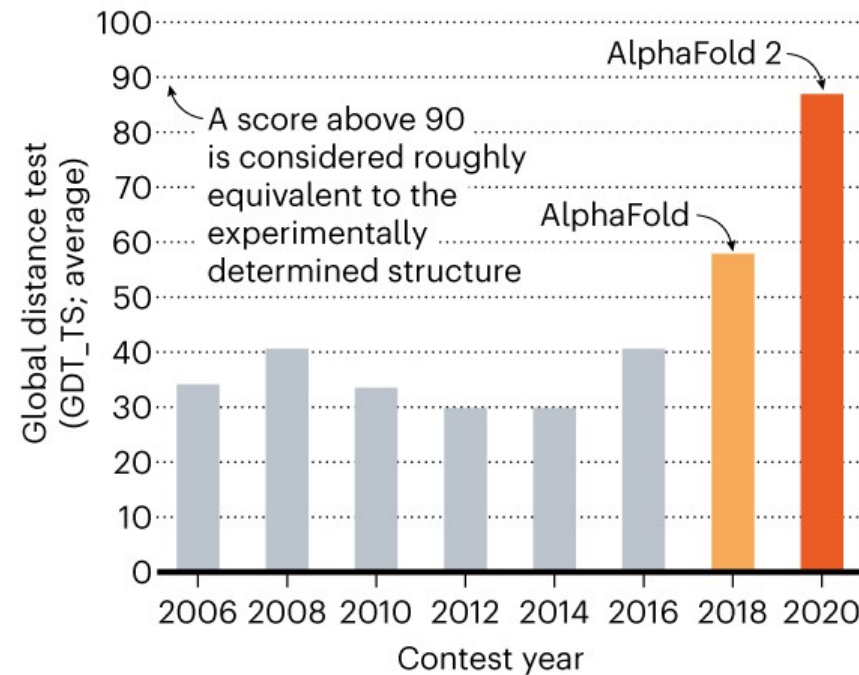
AlphaFold was a huge step forward

- DeepMind developed a deep learning approach to structure prediction for CASP13 (2016)
- Step-change in quality

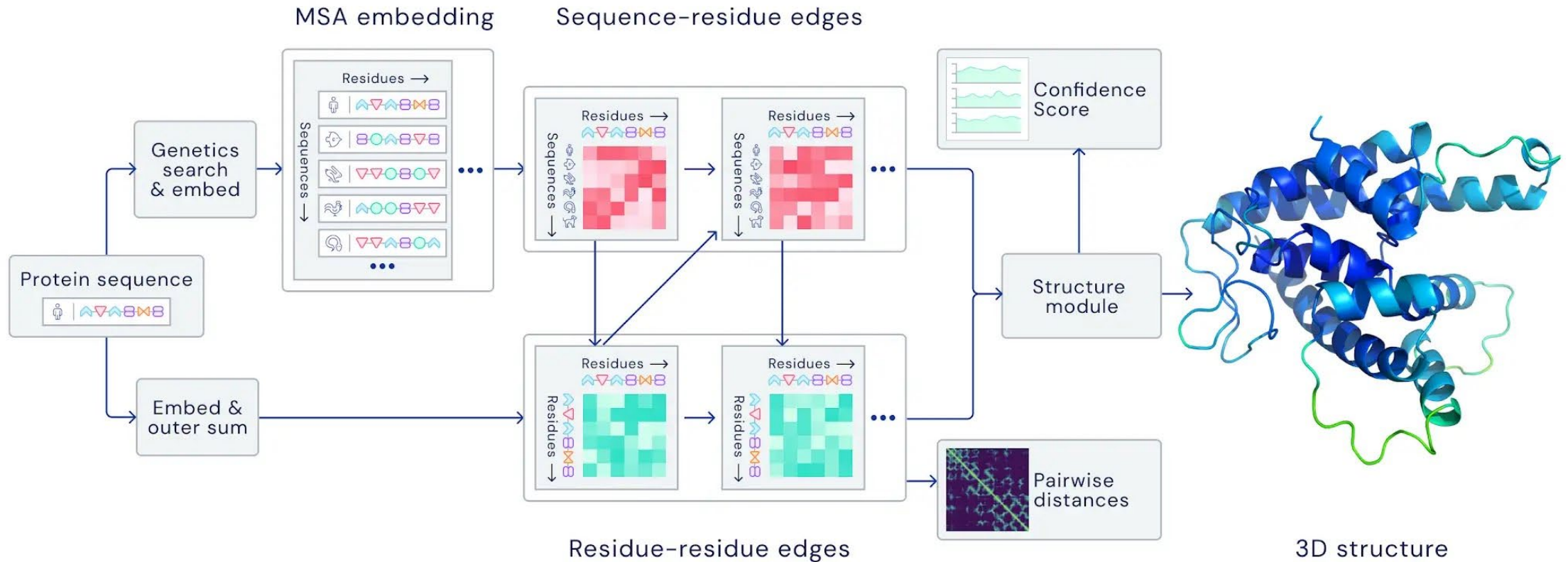


CASP14 and AlphaFold2

- DeepMind completely redesigned their prediction pipeline
- Unparalleled accuracy
- Quality of the prediction was largely decoupled from sequence identity to homologues

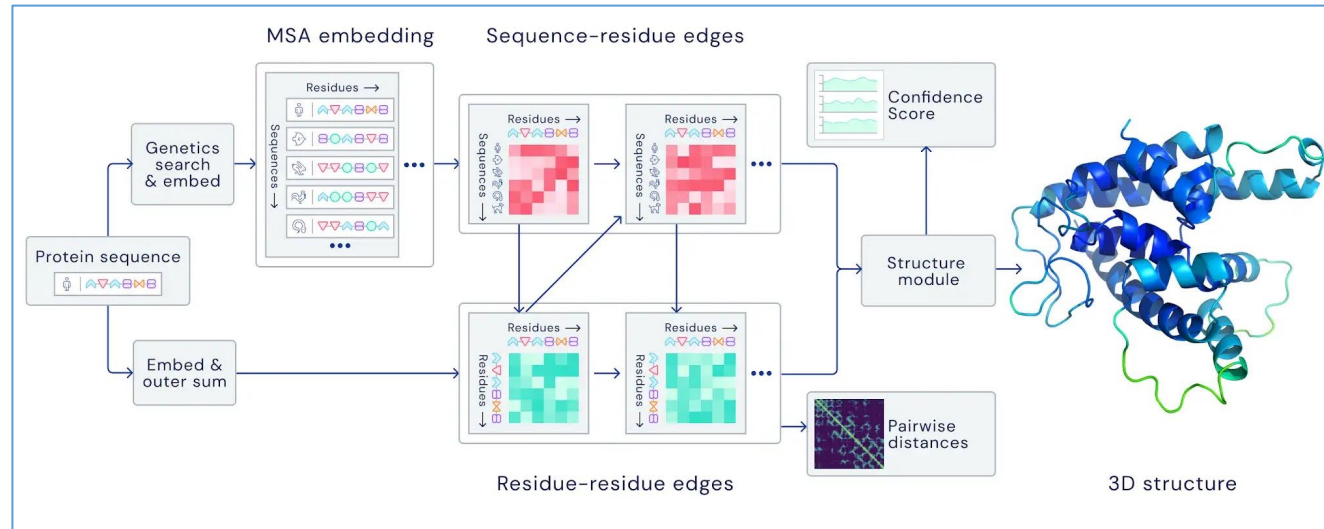


How does AlphaFold2 do this?



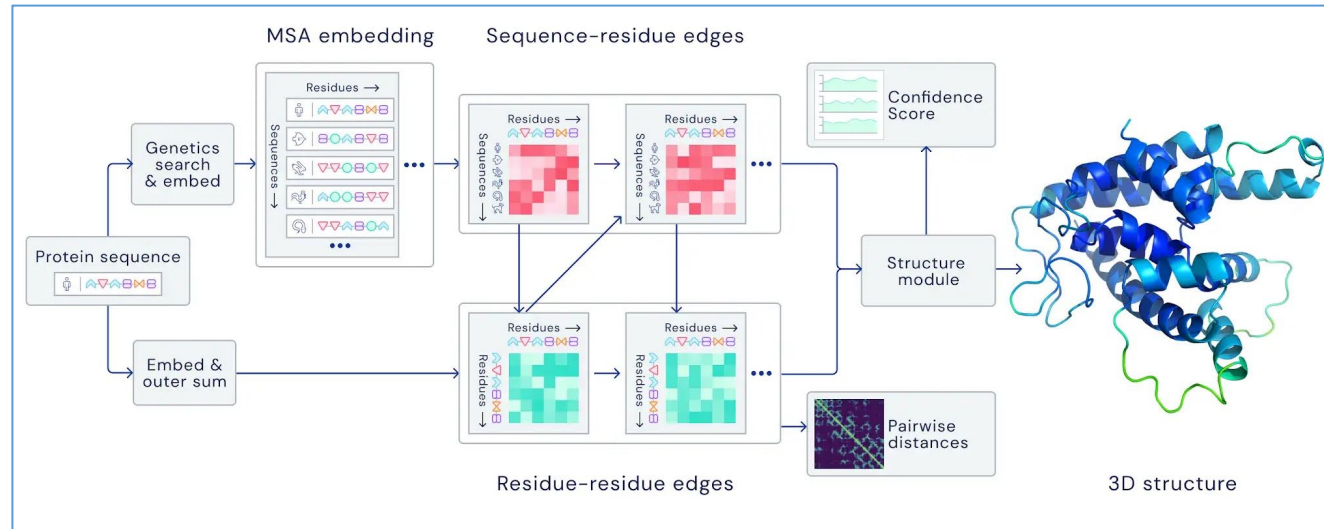
How does AlphaFold2 do this?

- The only input needed is a sequence! →



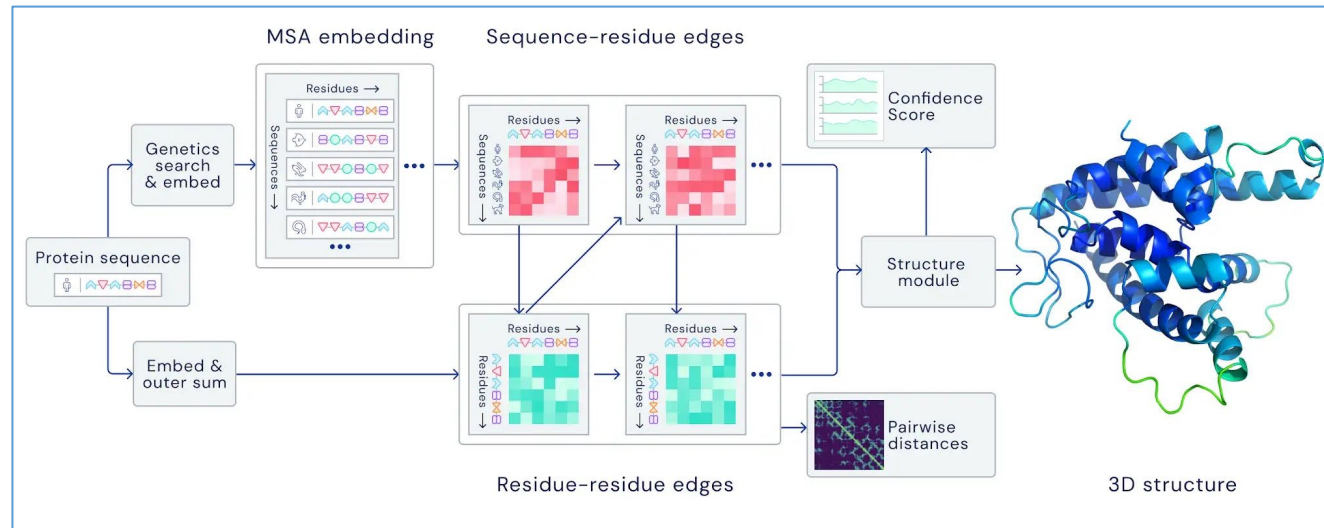
How does AlphaFold2 do this?

- Powerful neural networks
 - Attention-based neural networks



How does AlphaFold2 do this?

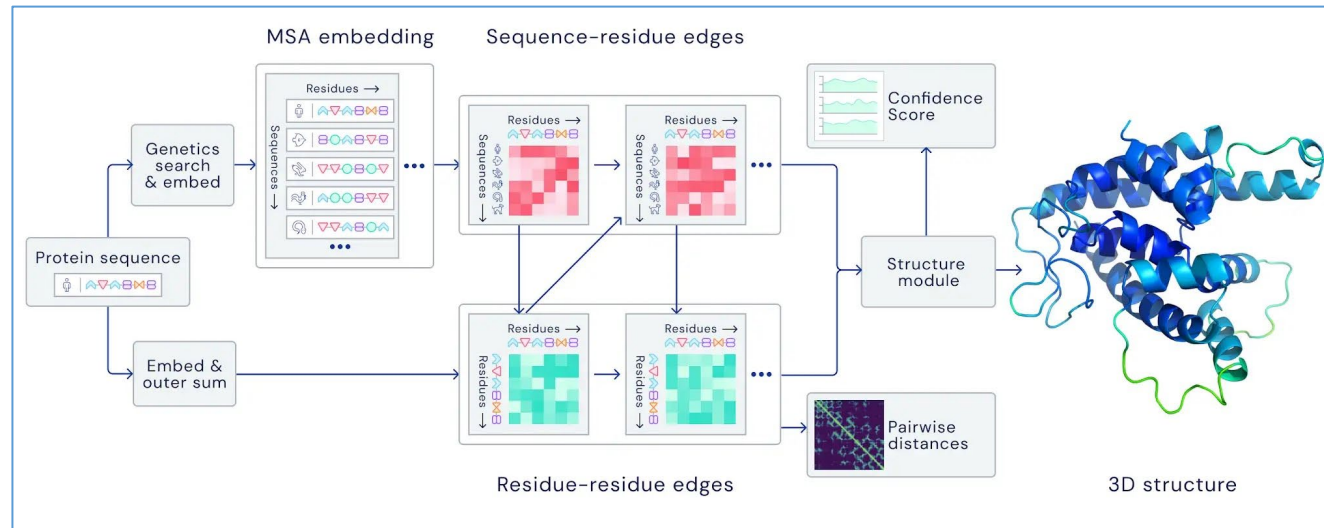
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- Protein sequences
 - Multiple sequence alignments
 - Interrogate co-evolution of residues

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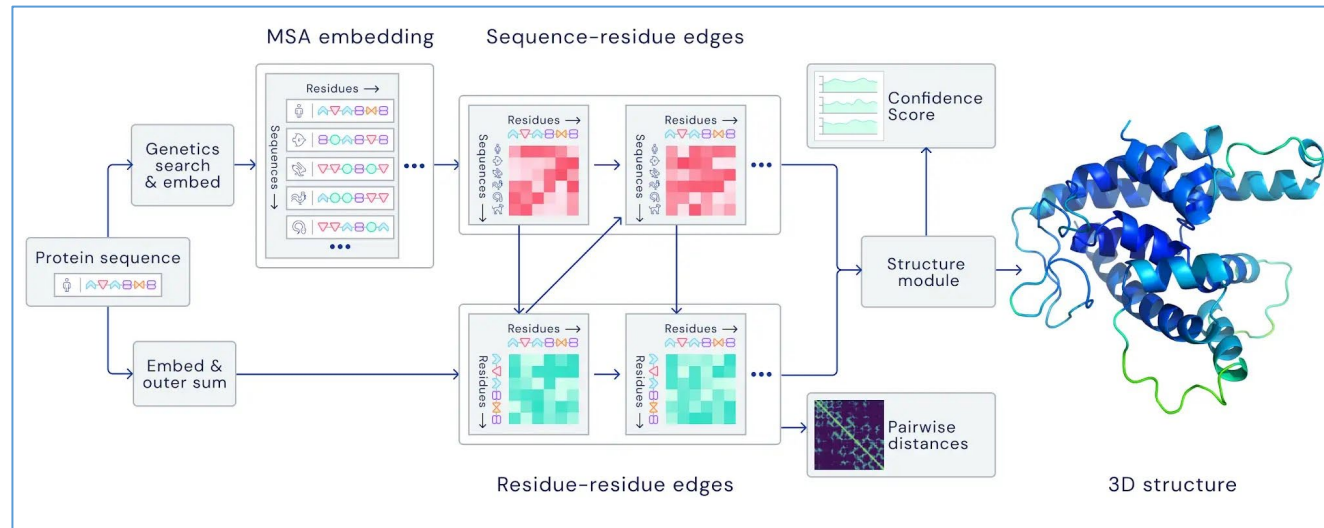


- Protein sequences
 - Multiple sequence alignments
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- Protein structures
 - Identifying residue pairs that should be close to each other
 - Experience of what folded proteins 'look like'

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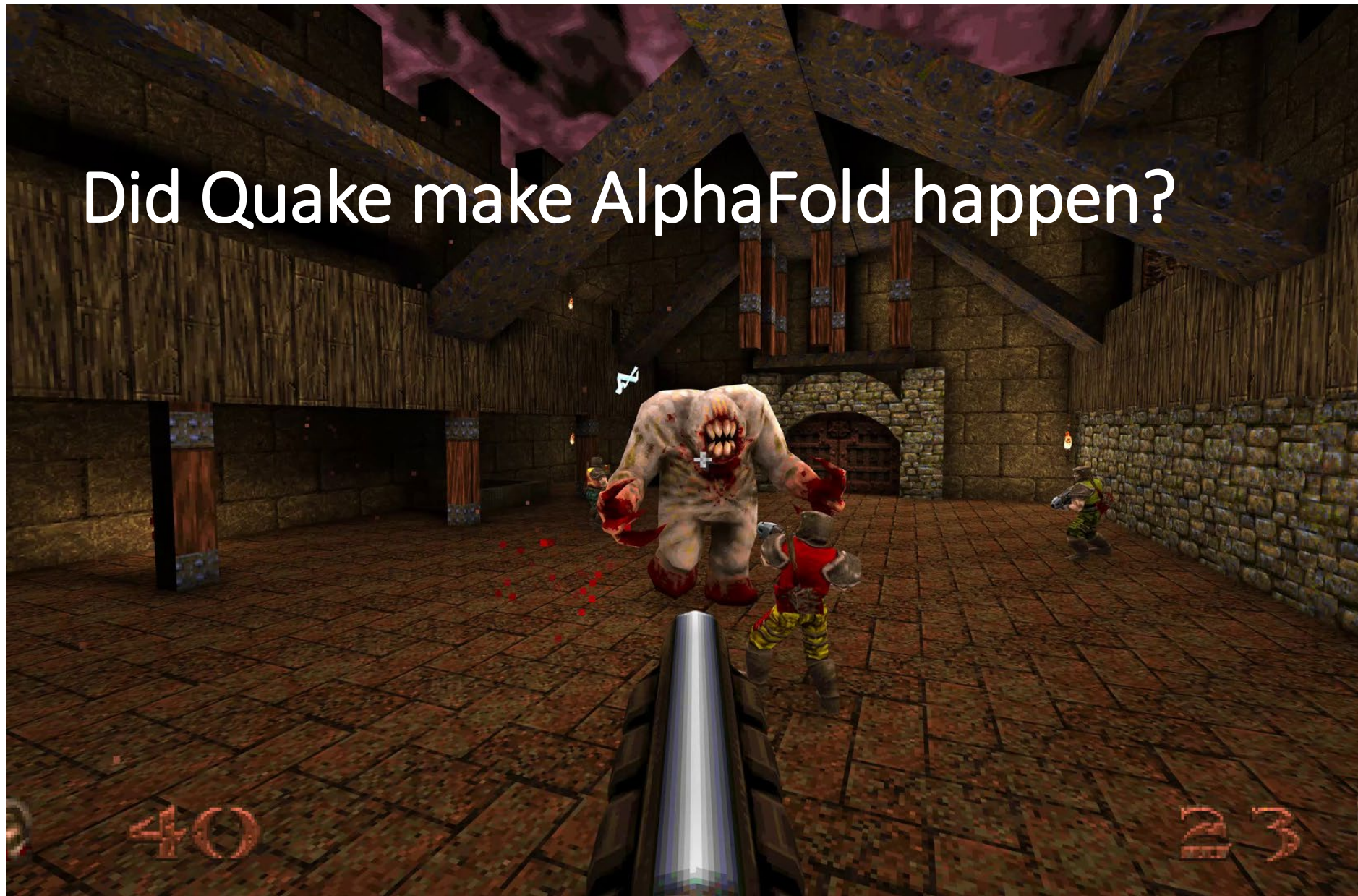


- Very powerful computers

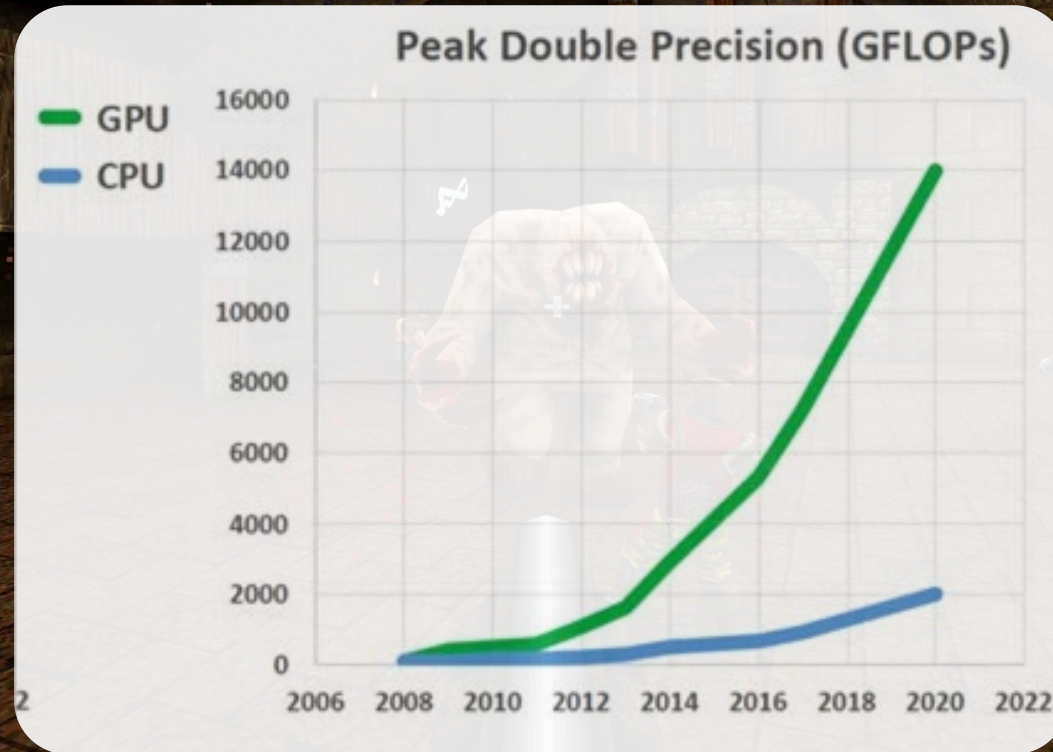
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Did Quake make AlphaFold happen?



Did Quake make AlphaFold happen?



AlphaFold2 in a nutshell

DeepMind

Highly accurate protein structure prediction with AlphaFold

John Jumper
AlphaFold lead, DeepMind

John Jumper^{1*}, Richard Evans^{1*}, Alexander Pritzel^{1*}, Tim Green^{1*}, Michael Figurnov^{1*}, Kathryn Tunyasuvunakool^{1*}, Olaf Ronneberger^{1*}, Russ Bates^{1*}, Augustin Židek^{1*}, Alex Bridgland^{1*}, Clemens Meyer^{1*}, Simon A A Kohl^{1*}, Anna Potapenko^{1*}, Andrew J Ballard^{1*}, Andrew Cowie^{1*}, Bernardino Romera-Paredes^{1*}, Stanislav Nikolov^{1*}, Rishub Jain^{1*}, Jonas Adler¹, Trevor Back¹, Stig Petersen¹, David Reiman¹, Martin Steinegger², Michalina Pacholska¹, David Silver¹, Oriol Vinyals¹, Andrew W Senior¹, Koray Kavukcuoglu¹, Pushmeet Kohli¹, Demis Hassabis^{1*}†

¹DeepMind, London, UK, ²Seoul National University, South Korea
* Equal contribution
† Corresponding authors: John Jumper (jumper@deepmind.com), Demis Hassabis (dhcontact@deepmind.com)

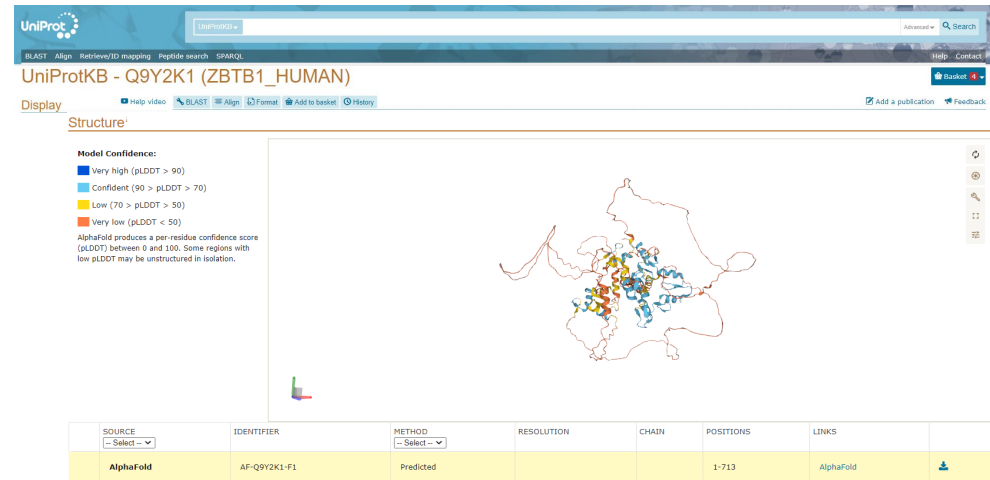
MRC LMB

- Kendrew Lecture 2021:
<https://www.youtube.com/watch?v=jT06odQNp90>



AF2 predictions of all proteins

- Teamed up with EBI to predict representative set of all known proteins (still ongoing...)
- Results for human and model organisms available already from Uniprot website



UniProtKB - Q9Y2K1 (ZBTB1_HUMAN)

Structure

Model Confidence:

- Very high (pLDDT > 90)
- Confident (90 > pLDDT > 70)
- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

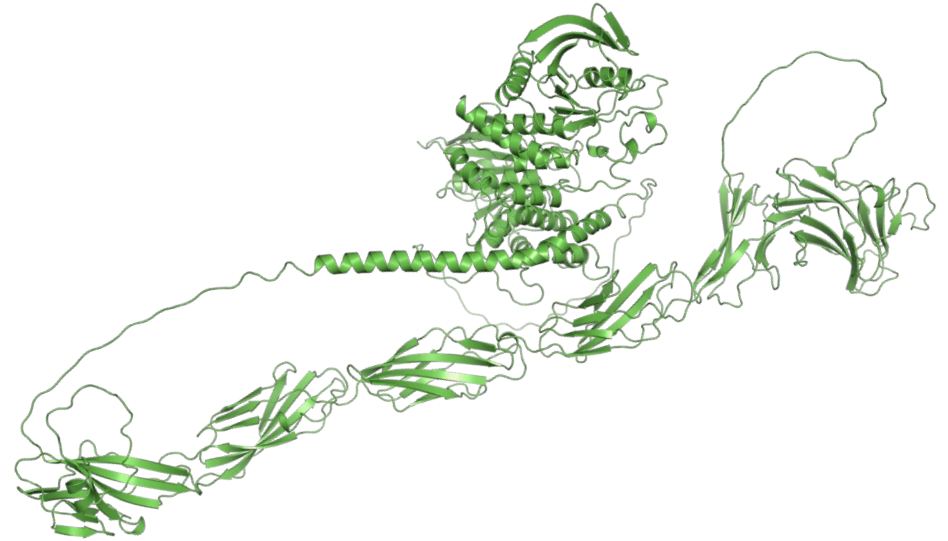
AlphaFold produces a per-residue confidence score (pLDDT) between 0 and 100. Some regions with low pLDDT may be unstructured in isolation.

SOURCE	IDENTIFIER	METHOD	RESOLUTION	CHAIN	POSITIONS	LINKS
AlphaFold	AF-Q9Y2K1-F1	Predicted			1-713	AlphaFold



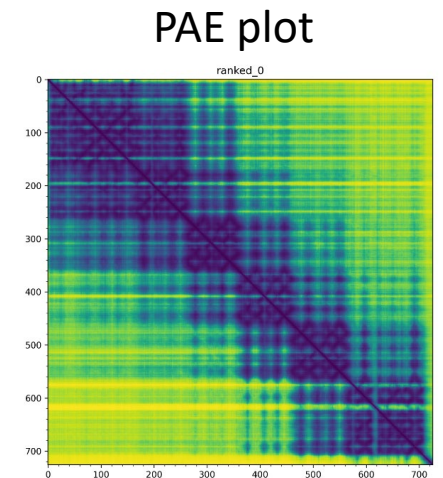
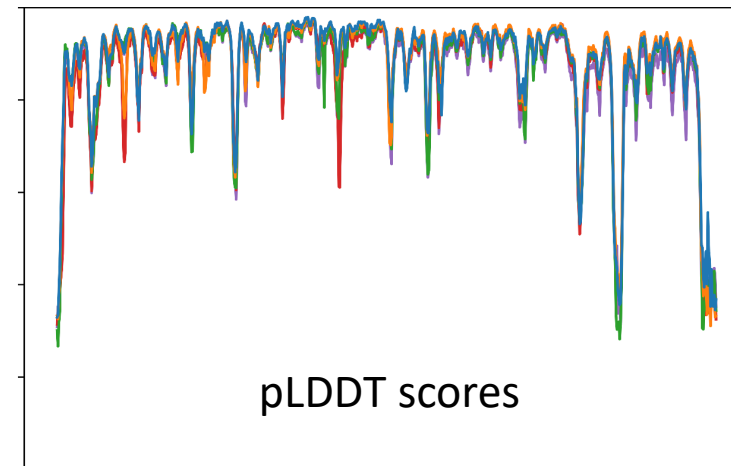
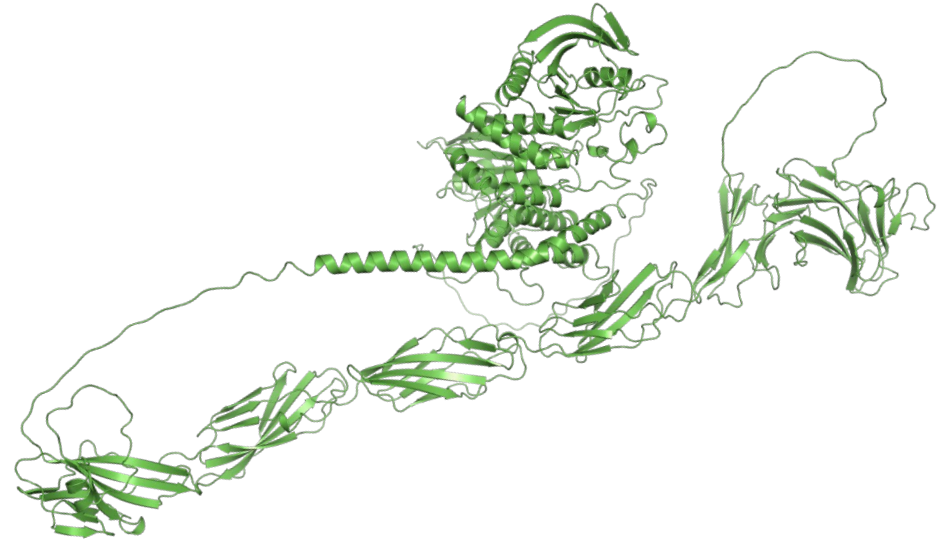
How reliable is your AF2 model?

- AF2 will always give you a structure
- But that doesn't mean it is right



How reliable is your AF2 model?

- AF2 will always give you a structure
- But that doesn't mean it is right
- **You have to check** the statistical plots and scores that are also generated



pLDDT Scores and Plot

- This is a per residue score on scale of 0 to 100
- Score above 70 is a confident prediction

Model Confidence:

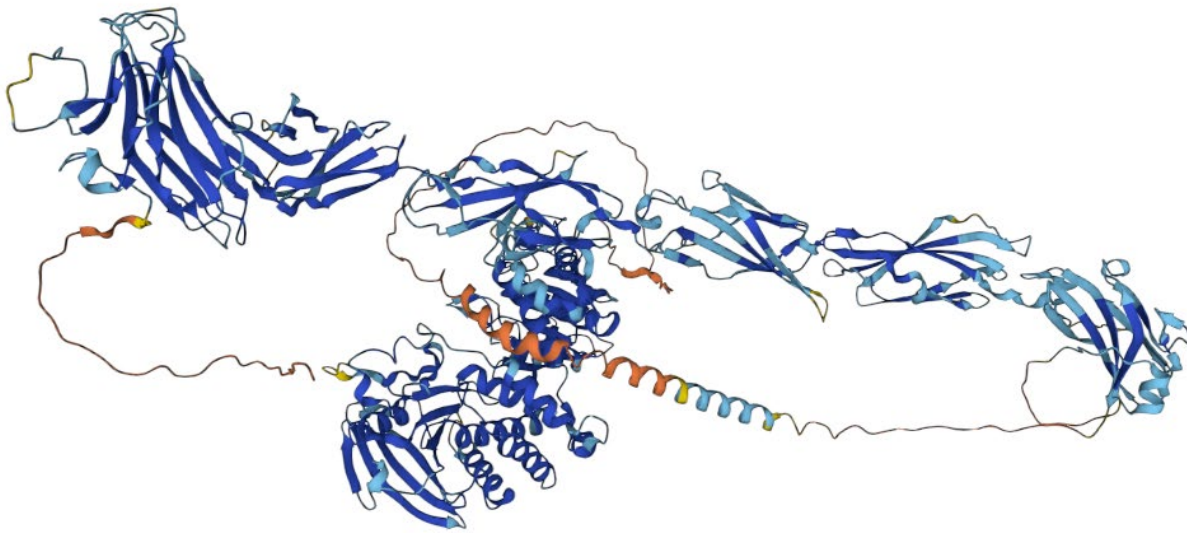
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pLDDT Scores and Plot

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pLDDT Scores and Plot

- This is a per residue score on scale of 0 to 100
- Score above 70 is a confident prediction
- Displayed as a coloured structure or a plot

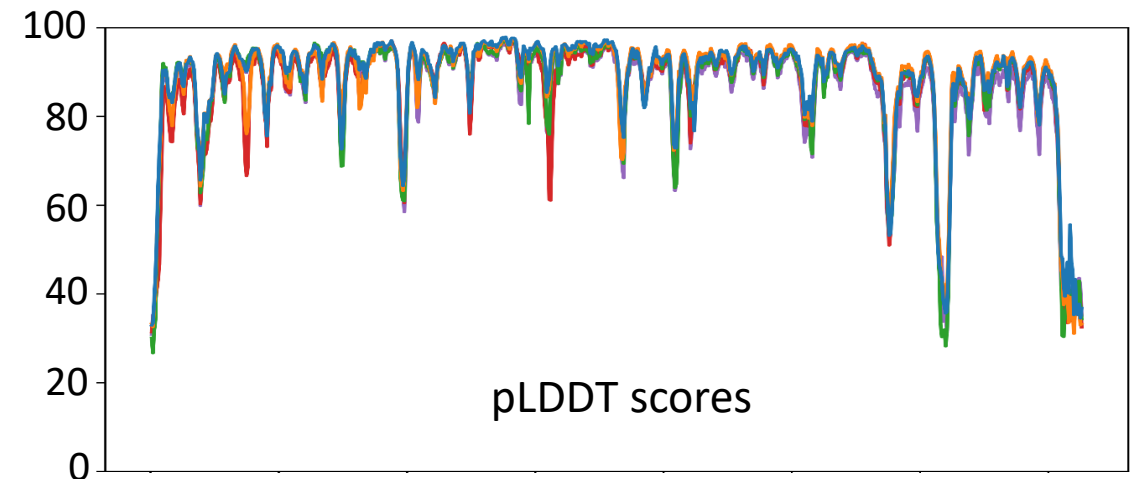
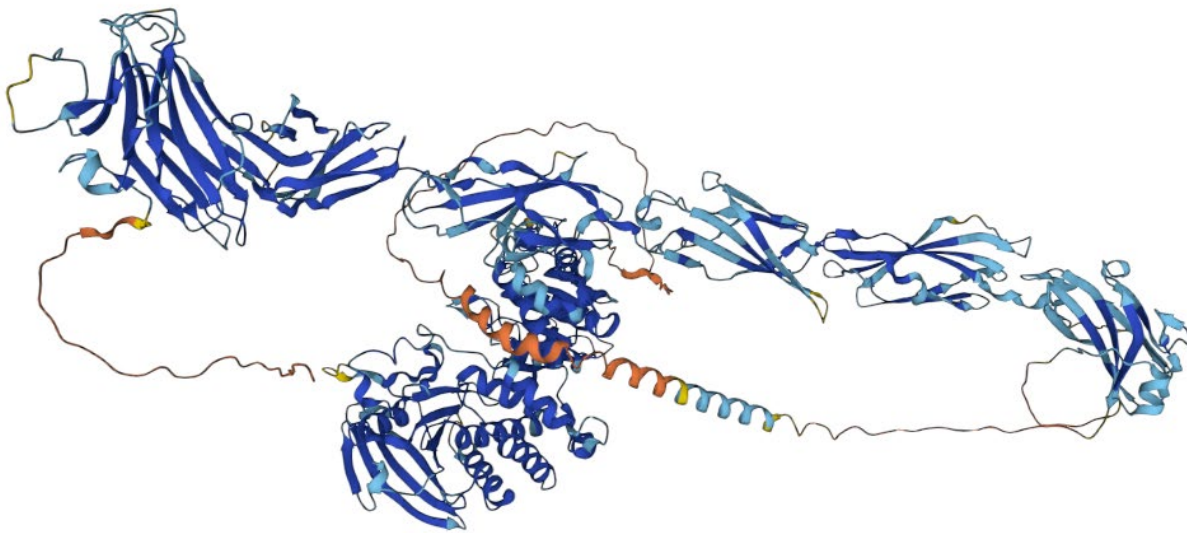
Model Confidence:

Very high (pLDDT > 90)

Confident (90 > pLDDT > 70)

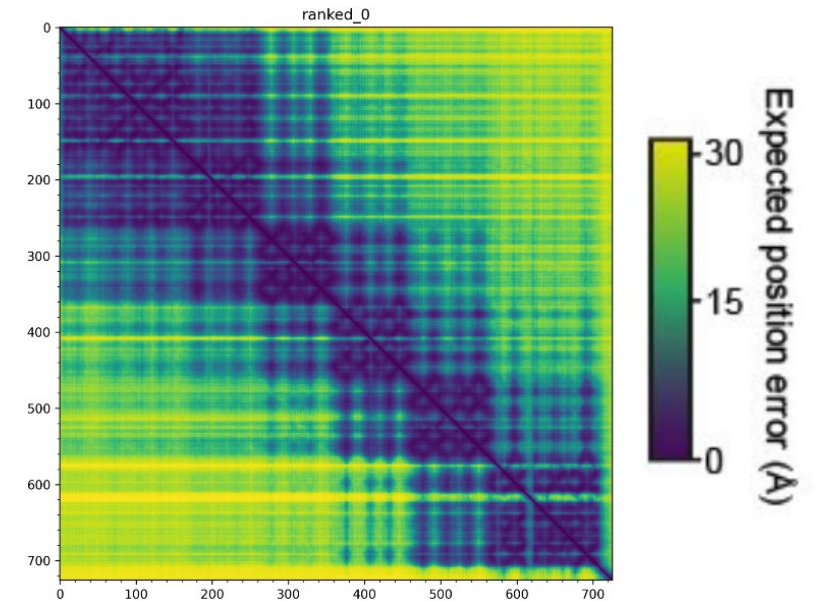
Low (70 > pLDDT > 50)

Very low (pLDDT < 50)



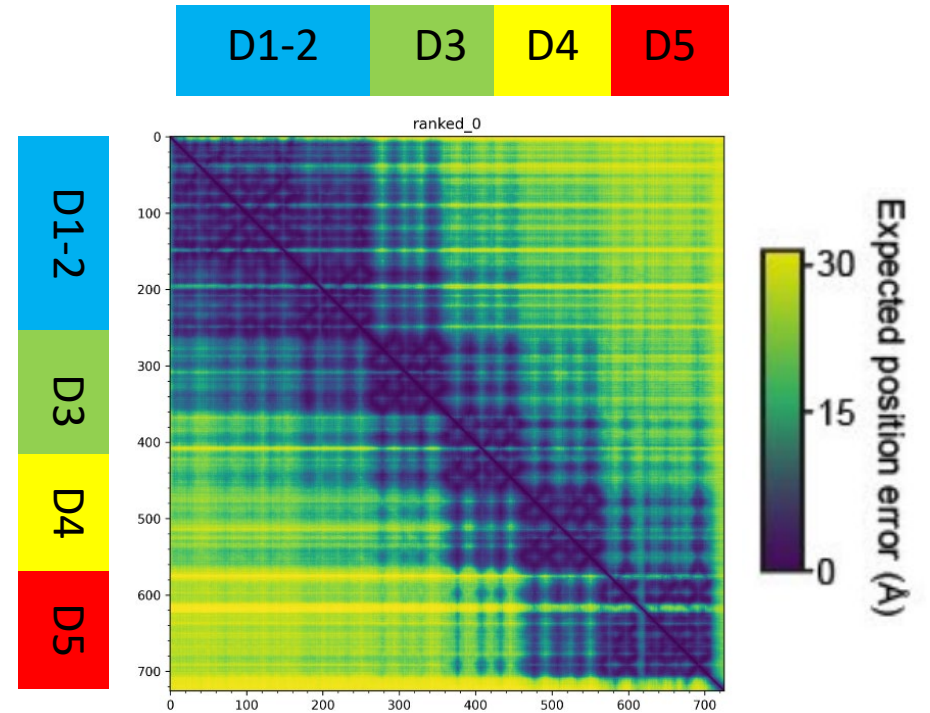
PAE plots – Predicted Aligned Error

- This is a measure of confidence relative to other regions of the structure
- Low error is high confidence (blue)
- High error is low confidence (yellow)

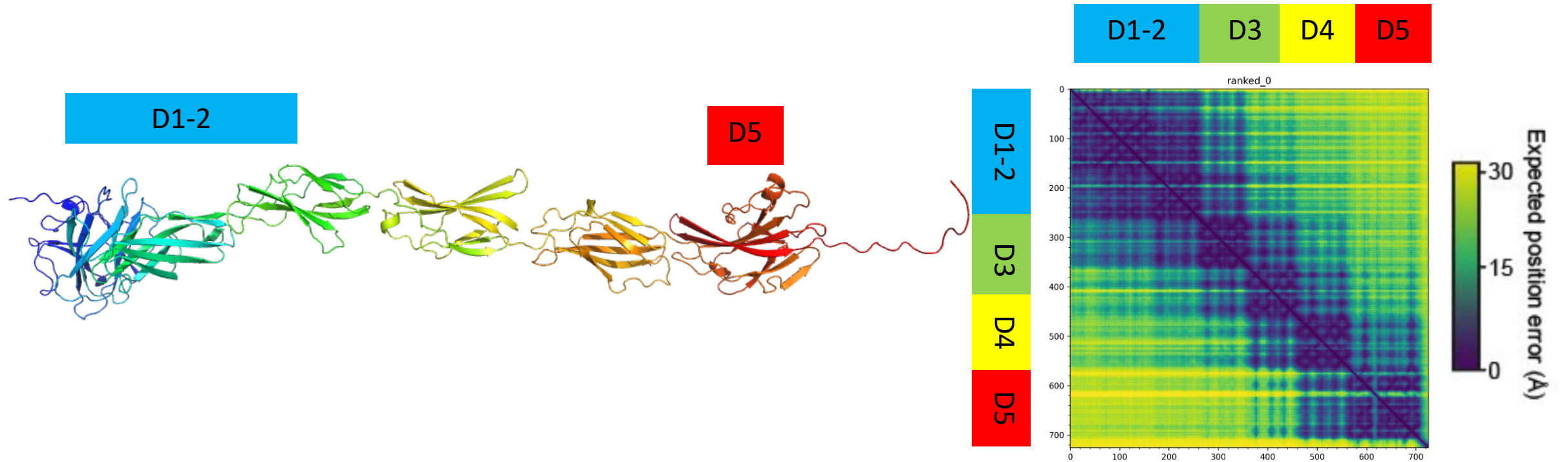


PAE plots – Predicted Aligned Error

- This is a measure of confidence relative to other regions of the structure
- Low error is high confidence (blue)
- High error is low confidence (yellow)
- In this example:
 - Domains 1 and 2 confident relative to each other
 - Domains 1 and 2 not confident relative to domains 4 and 5

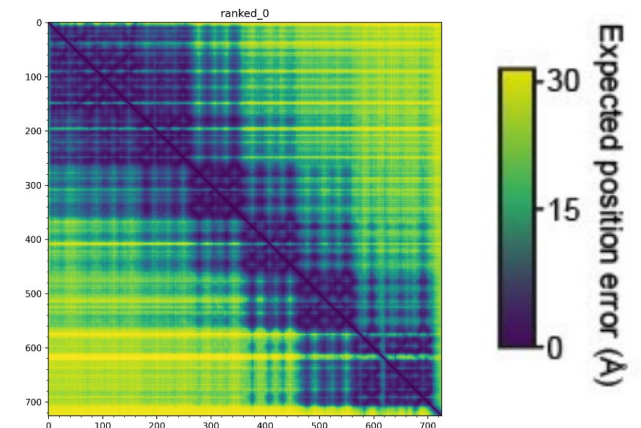
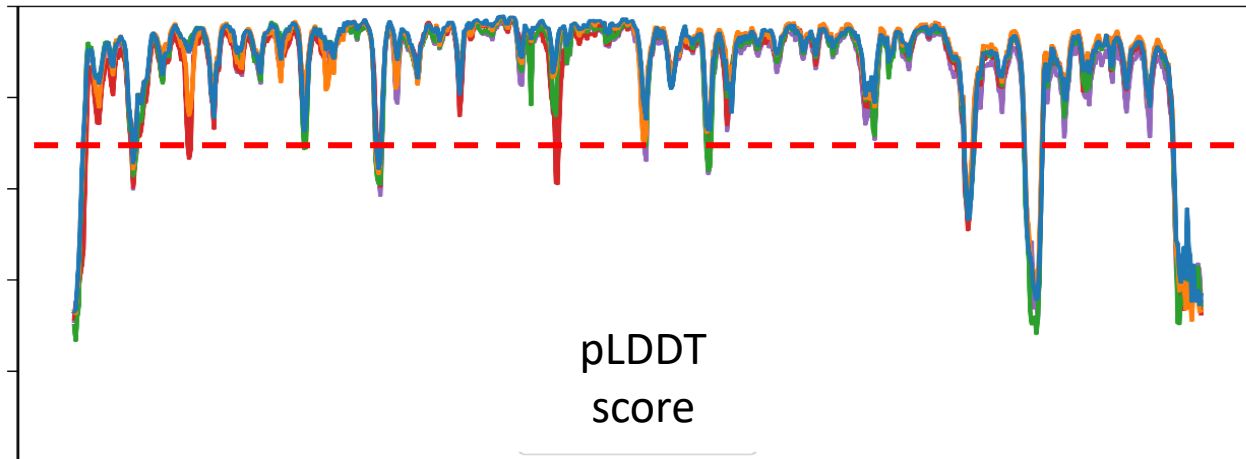
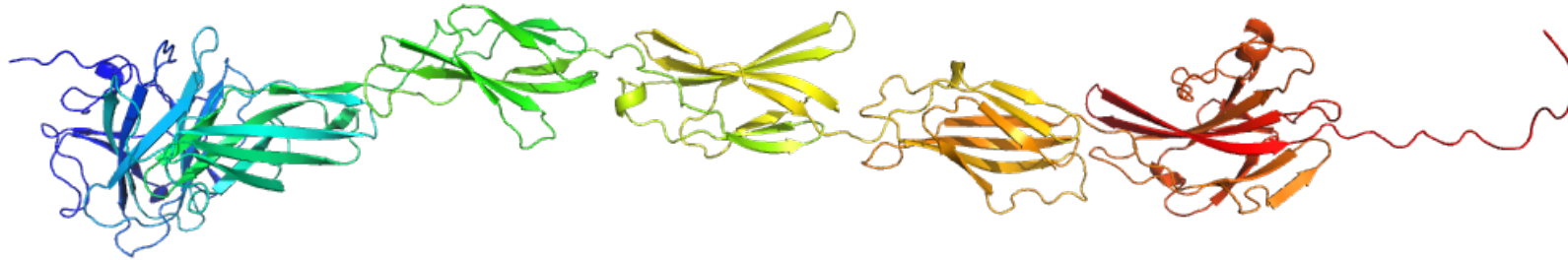


PAE plots – Predicted Aligned Error



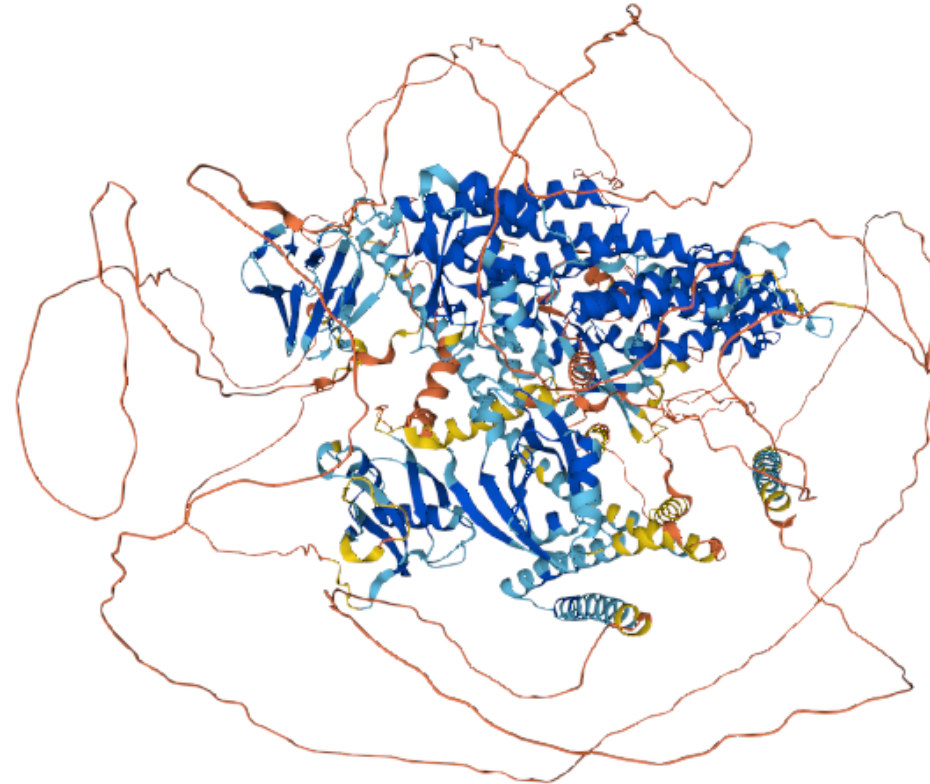
How to judge an AF2 model

- To summarise, a high-confidence per-residue model can be low-confidence overall



What about proteins like this?

Human
Afadin

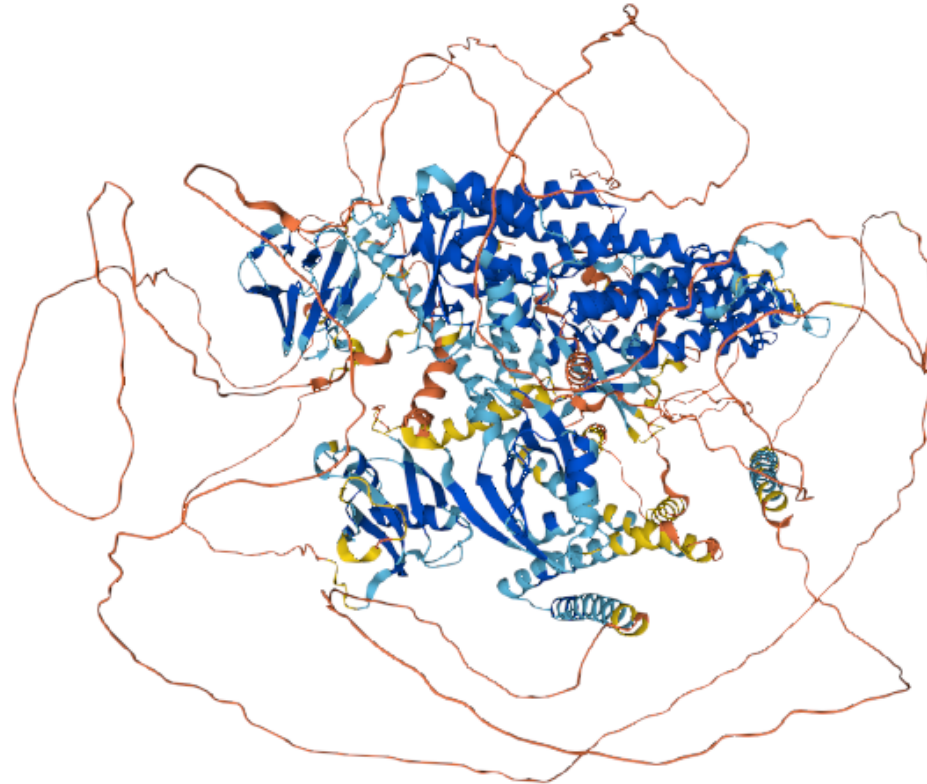


Predicted
accuracy
High → Low

- If you see something like this, can you learn anything at all?

What about proteins like this?

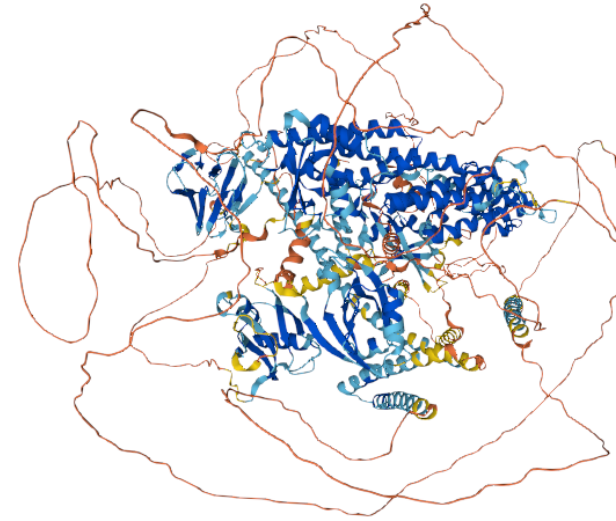
Human
Afadin



Predicted
accuracy
High → **Low**

Actually quite high
confidence these
regions are
unstructured

Look at the pLDDT plot

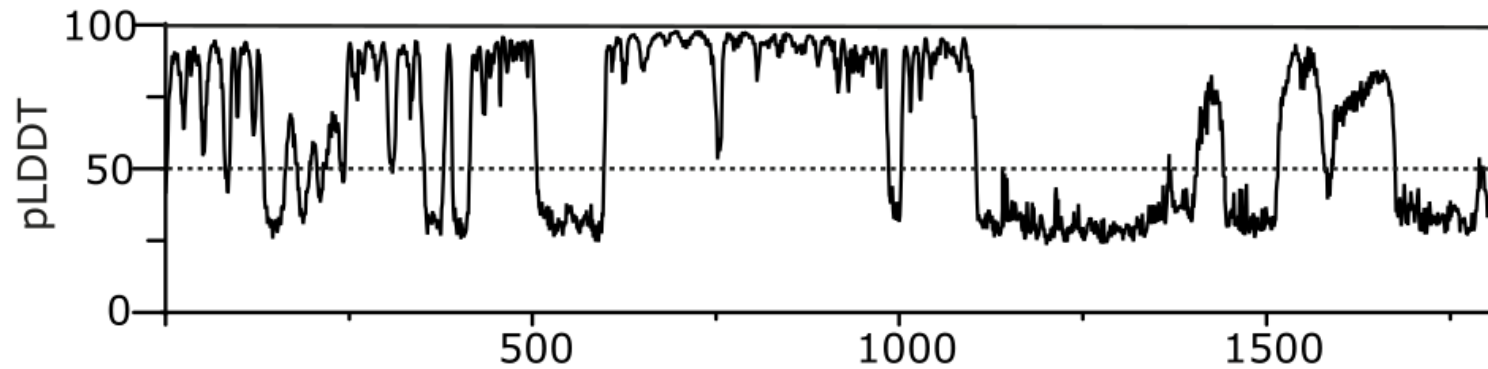


Predicted accuracy

High

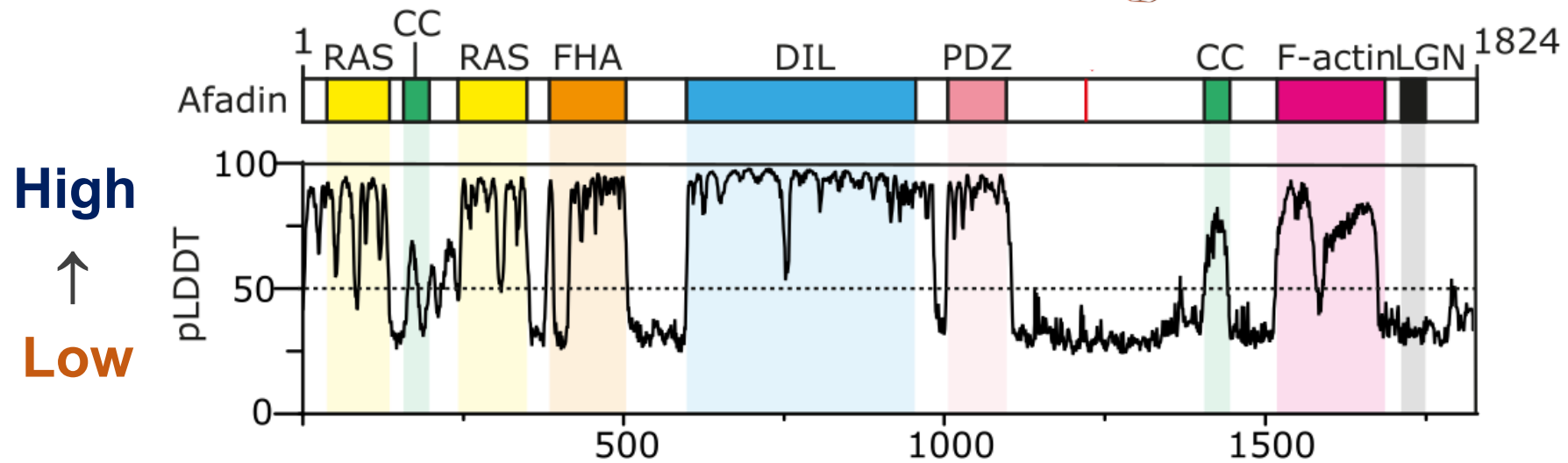
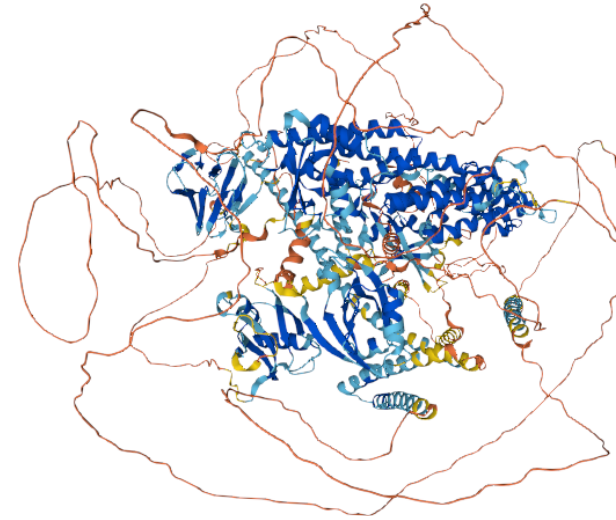


Low

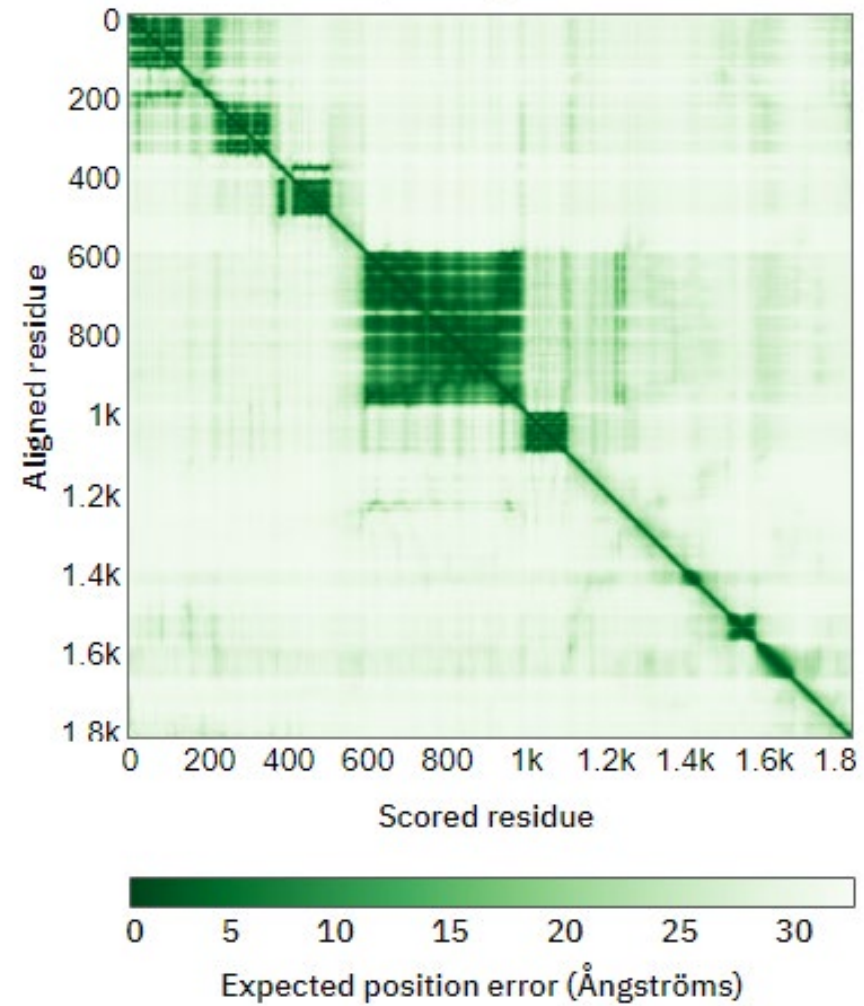
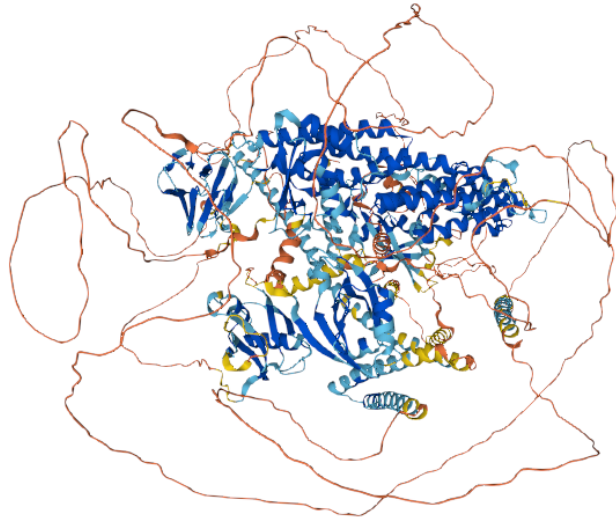


Look at the pLDDT plot

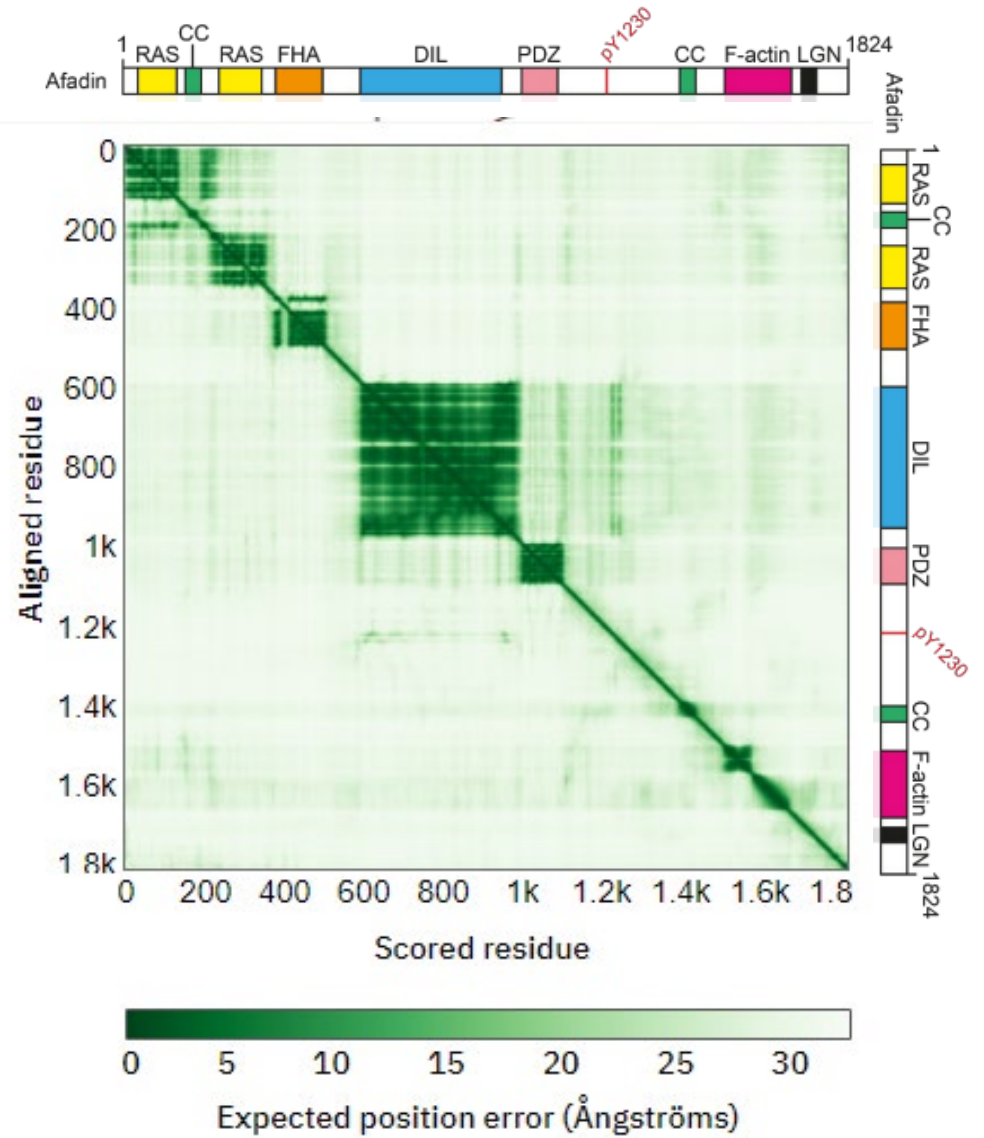
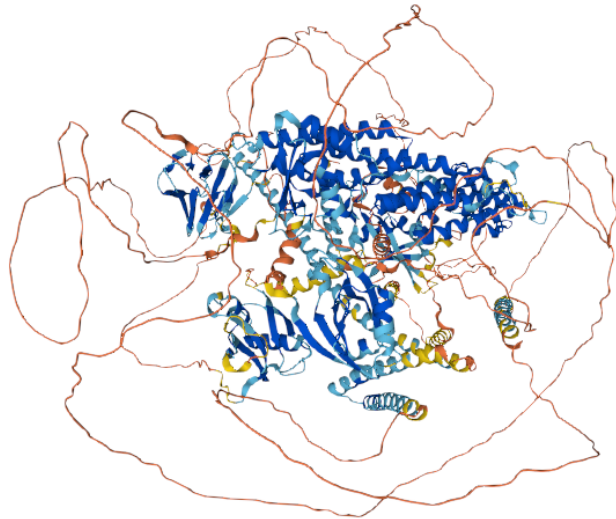
- Clearly determine domain boundaries



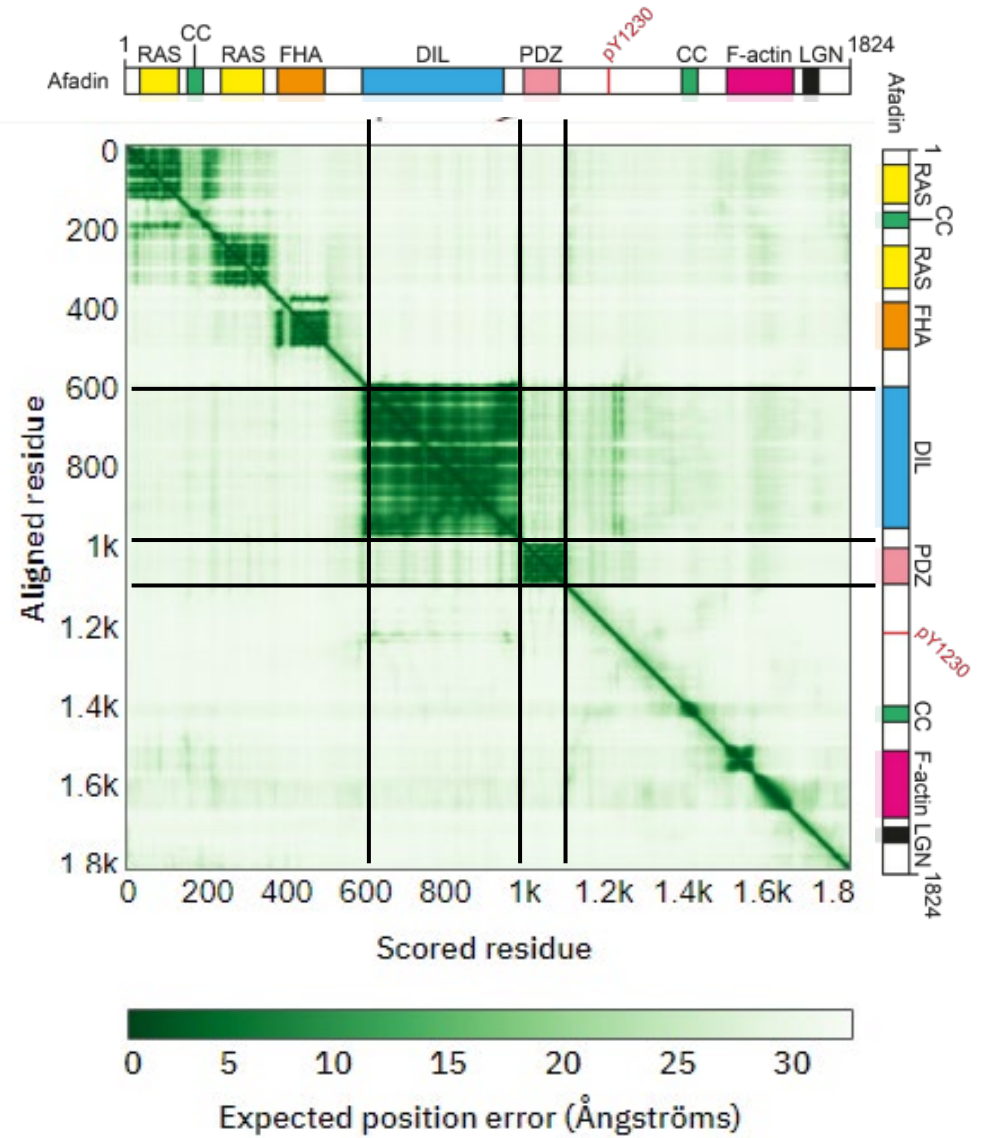
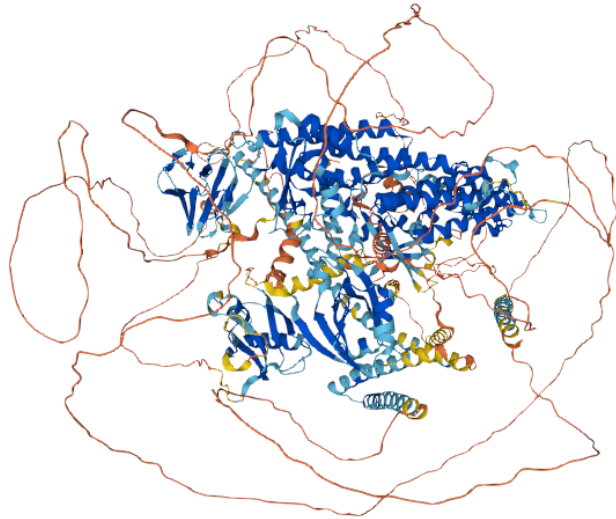
Look at the PAE plot



Look at the PAE plot

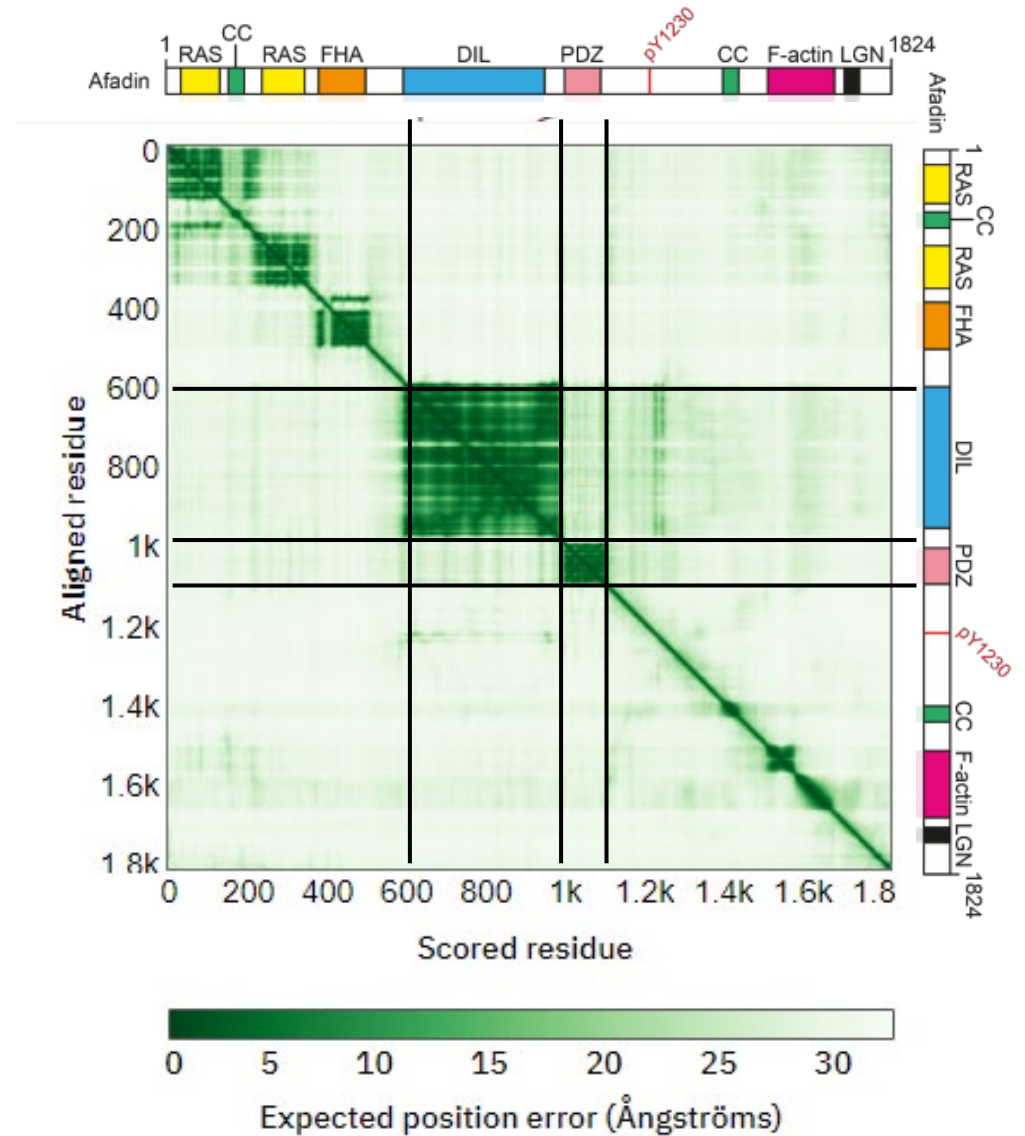


Look at the PAE plot



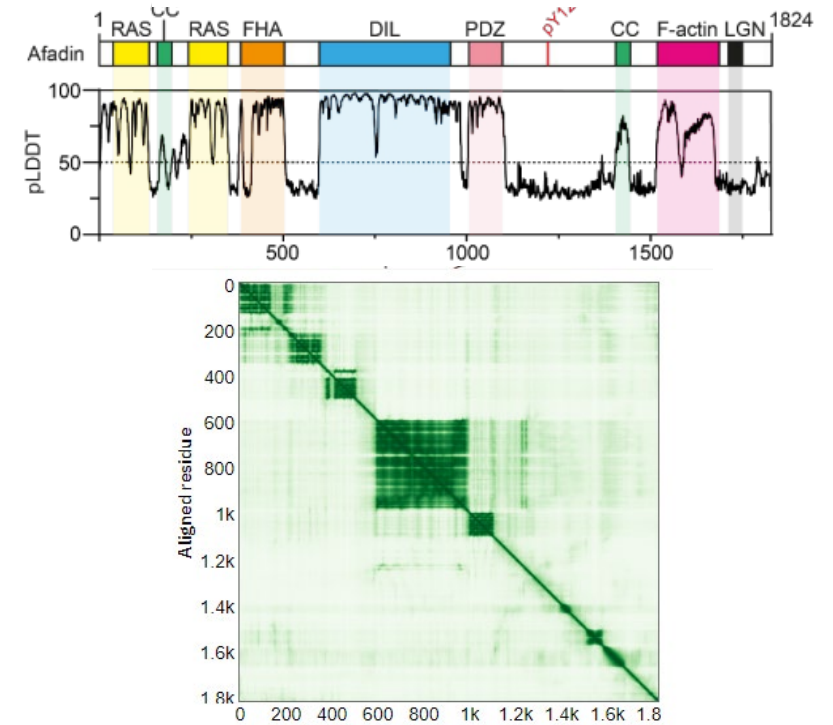
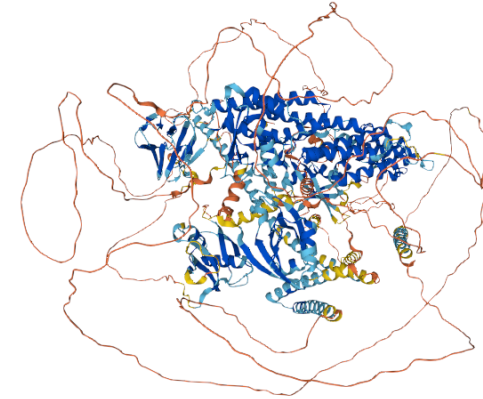
Look at the PAE plot

- The predictions are only confident within domains
NOT BETWEEN



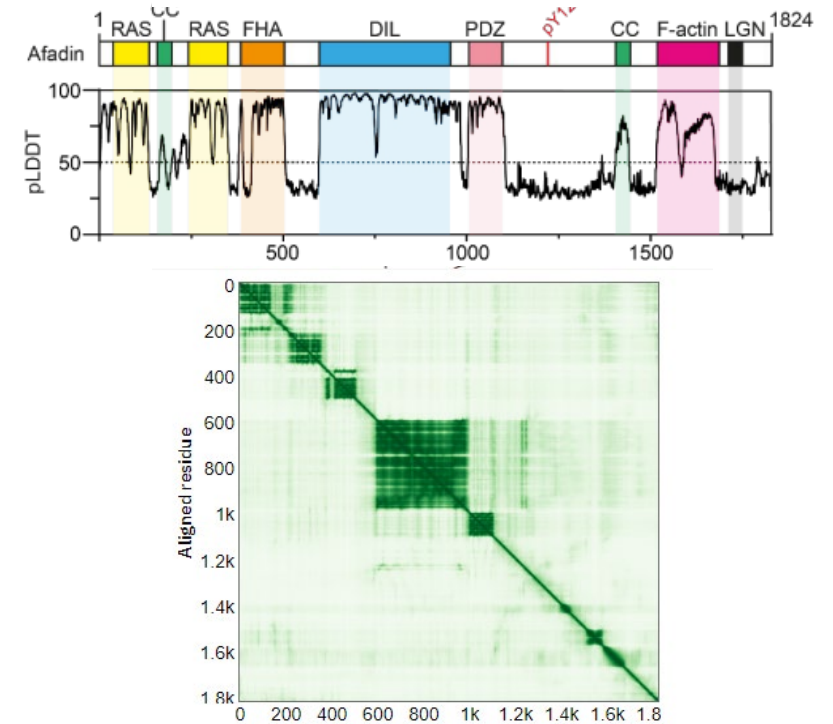
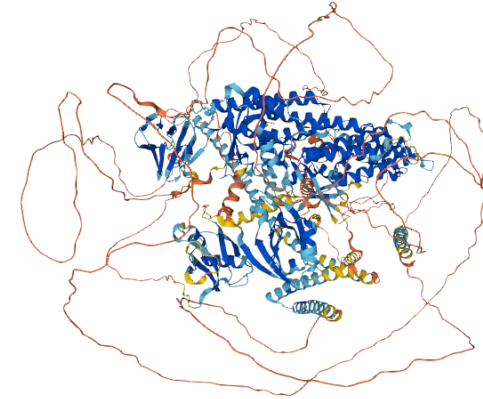
In this case:

- This AF2 model is useful for:
 - Determining domain boundaries
 - Fold of individual domains

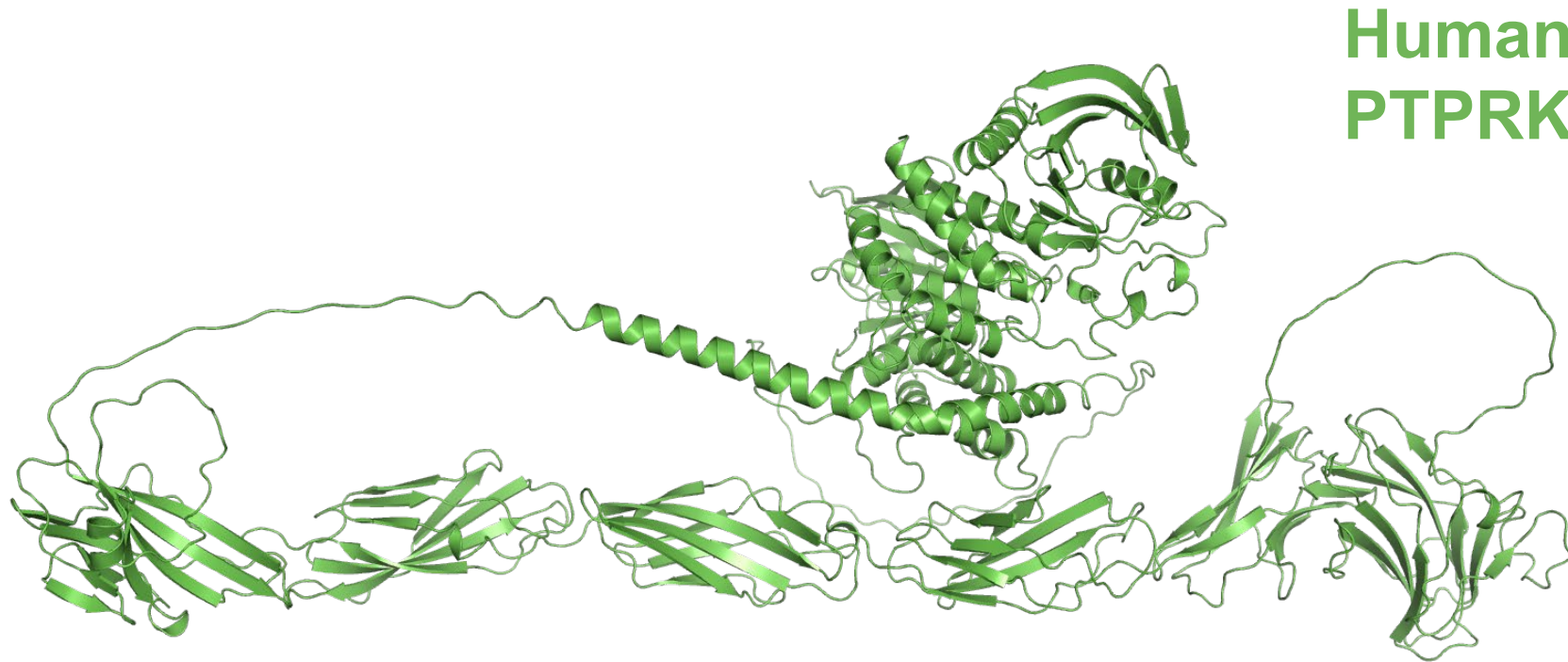


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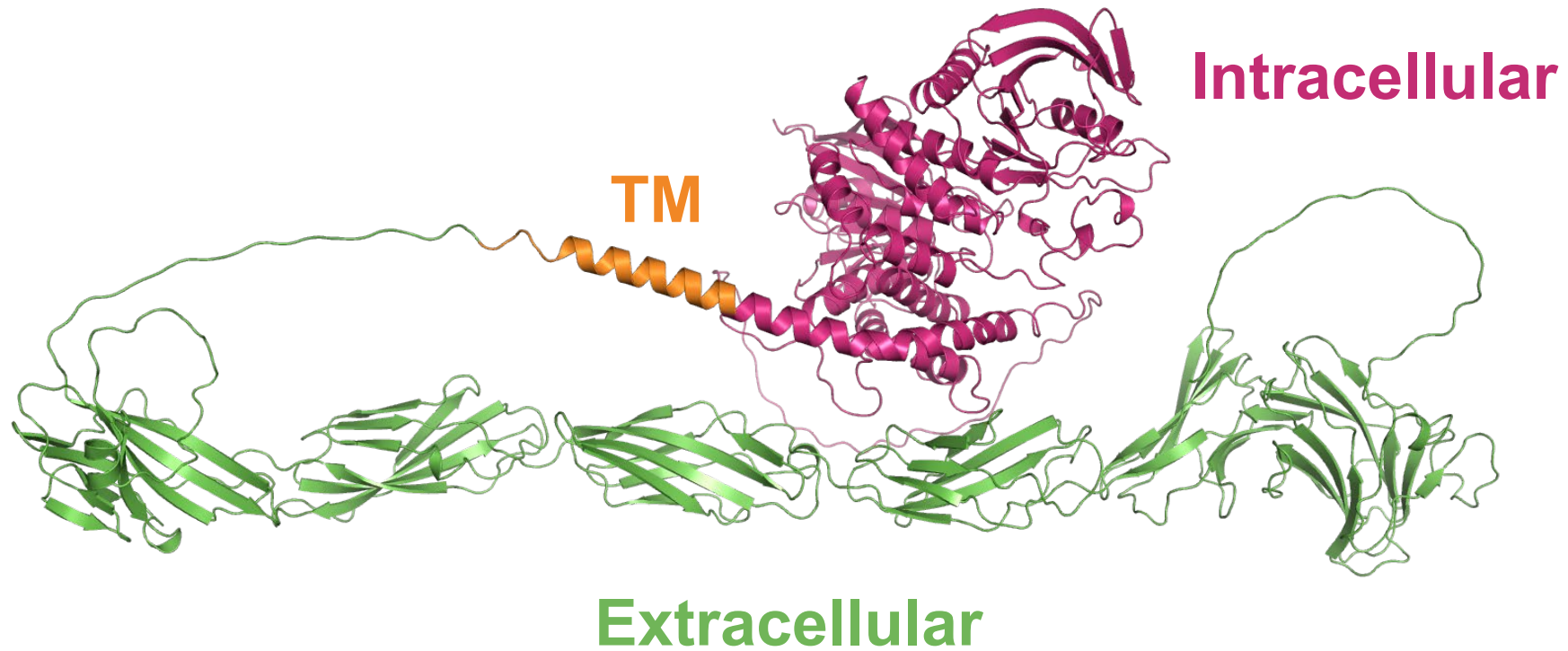
- This AF2 model is useful for:
 - Determining domain boundaries
 - Fold of individual domains
 - You could then use these individual domains to search using DALI or FoldSeek to find structural homologues that may inform function



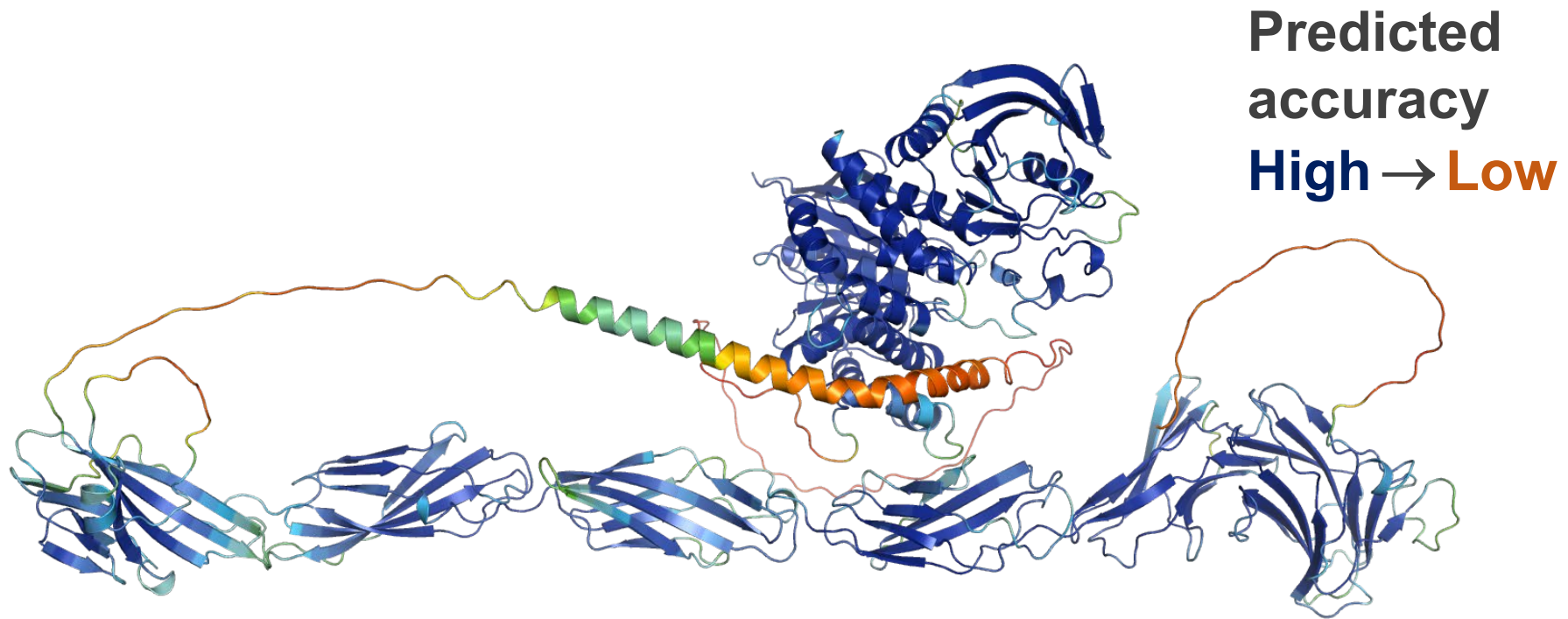
AF2 doesn't know about topology



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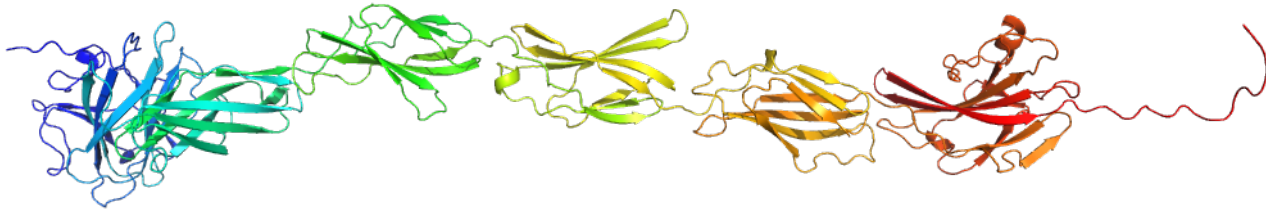


- Treat membrane-spanning models with caution...



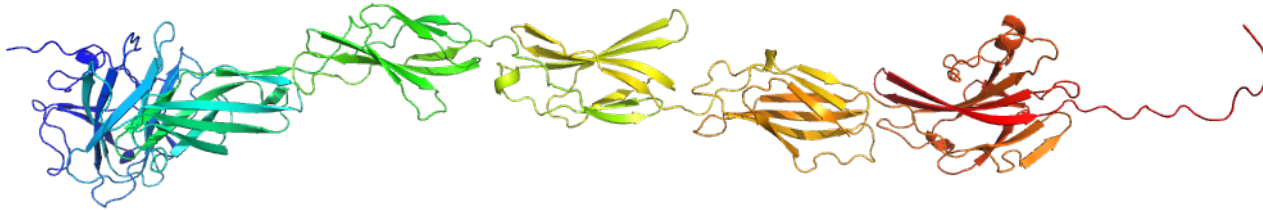
Validate using experimental techniques

AlphaFold Prediction

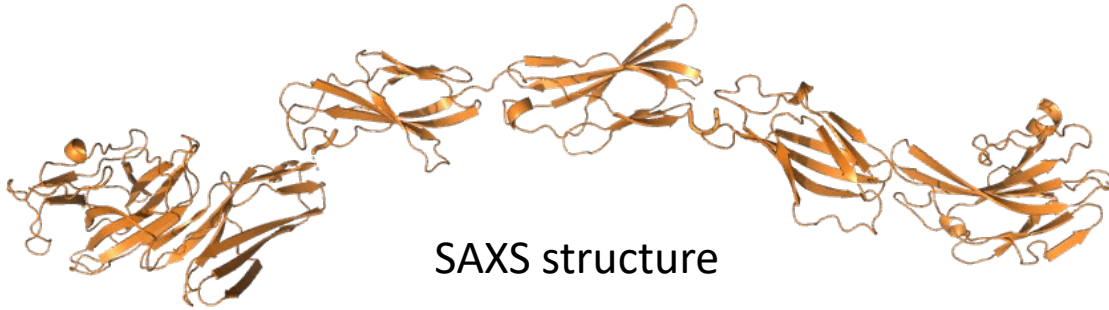


Validate using experimental techniques

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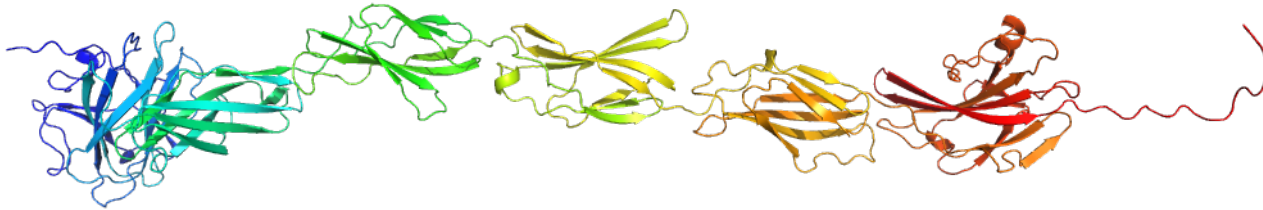


SAXS structure

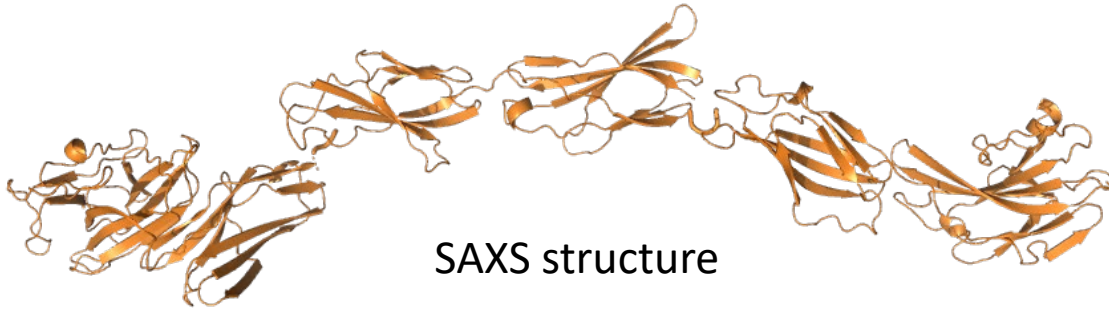


Validate using experimental techniques

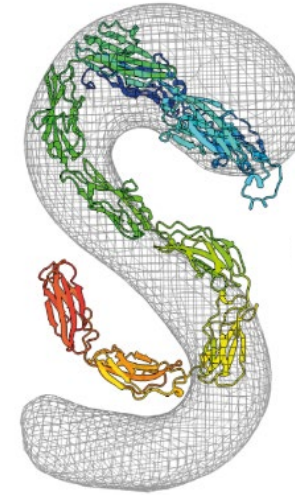
AlphaFold Prediction



SAXS structure

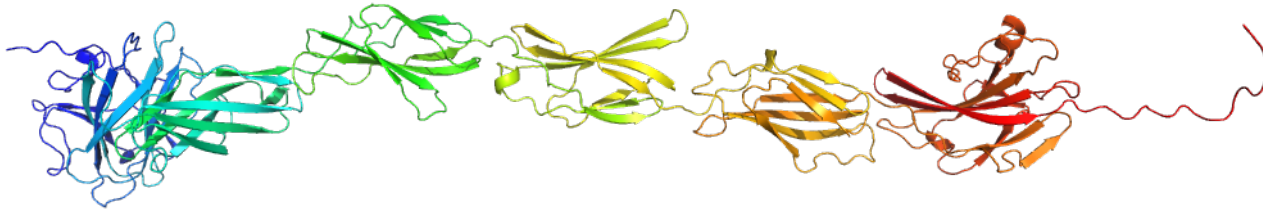


AlphaFold Prediction

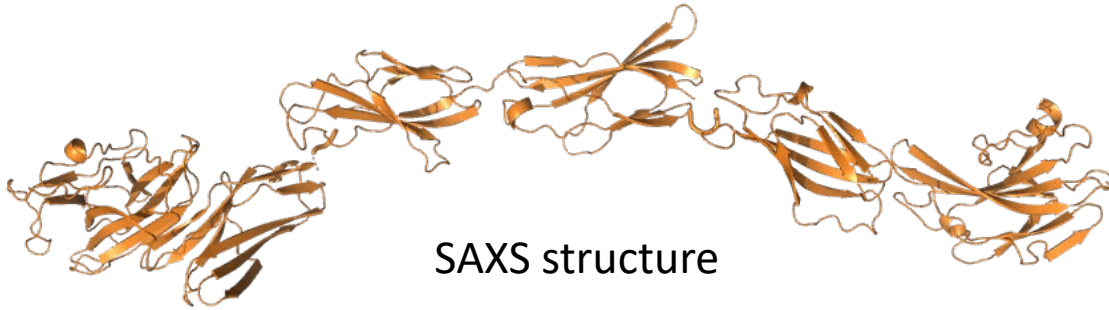


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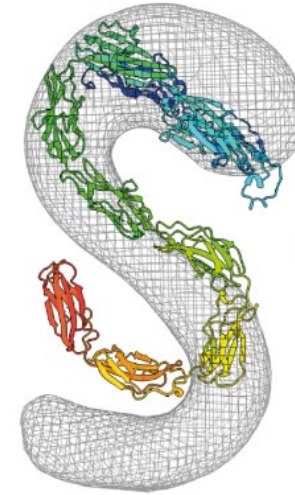
AlphaFold Prediction



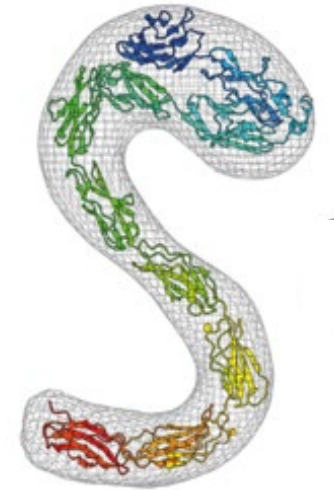
SAXS structure



AlphaFold Prediction

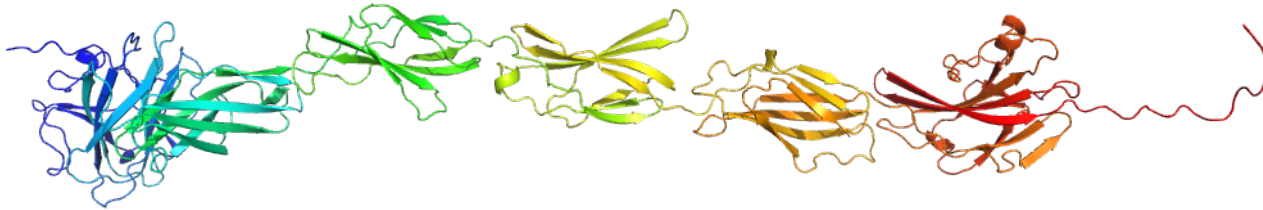


EM structure

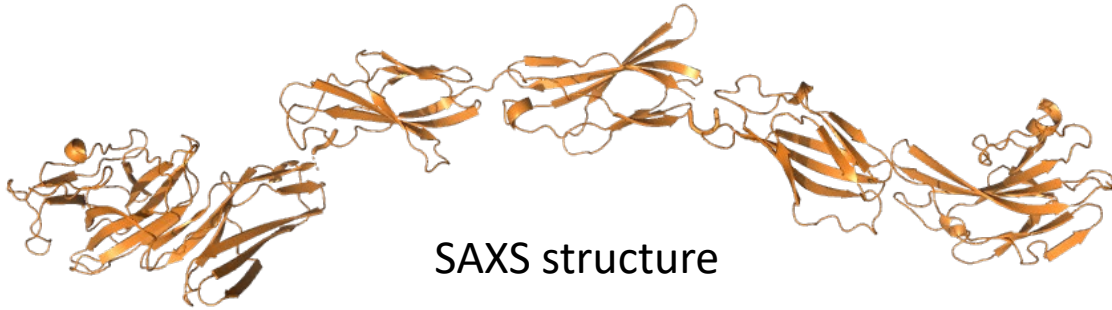


Validate using experimental techniques

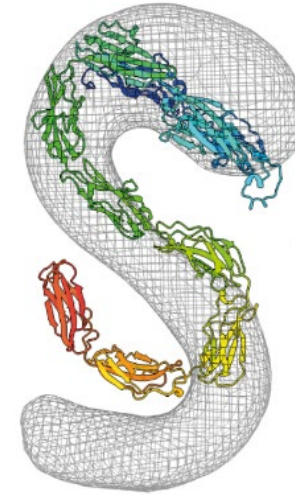
AlphaFold Prediction



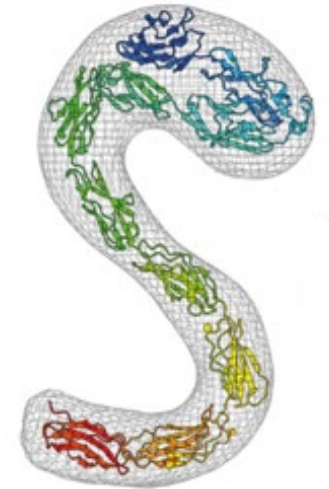
SAXS structure



AlphaFold Prediction



EM structure



- Importantly, AF2 provided excellent starting models for these experimental approaches

AF2 is pretty good at single proteins...what about complexes?

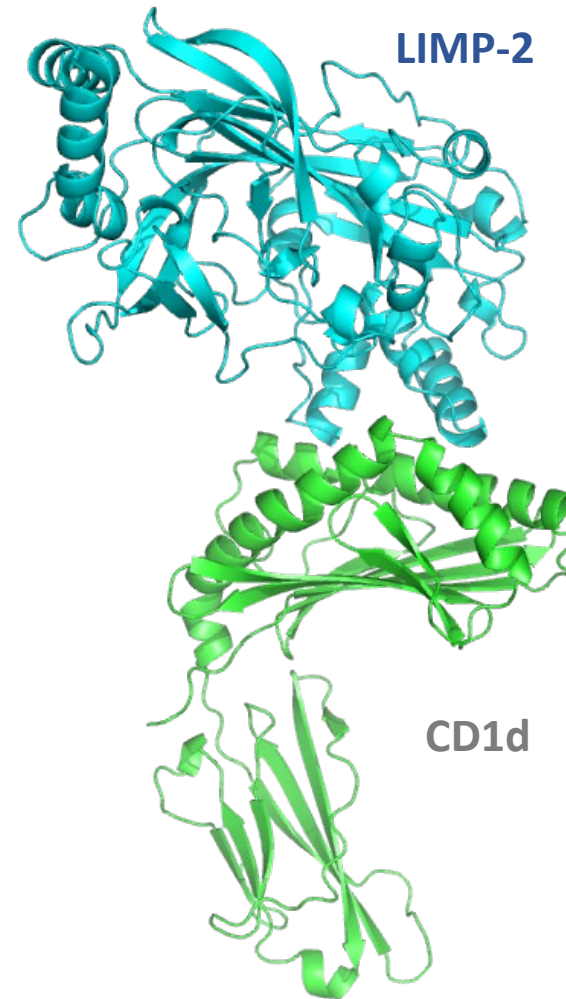
- AF2 Multimer was developed to try and address this question
- Answer is mixed, again you have to know how to interpret the statistics of the models produced
- Two examples: CD1d-LIMP2

PTPRK-Afadin



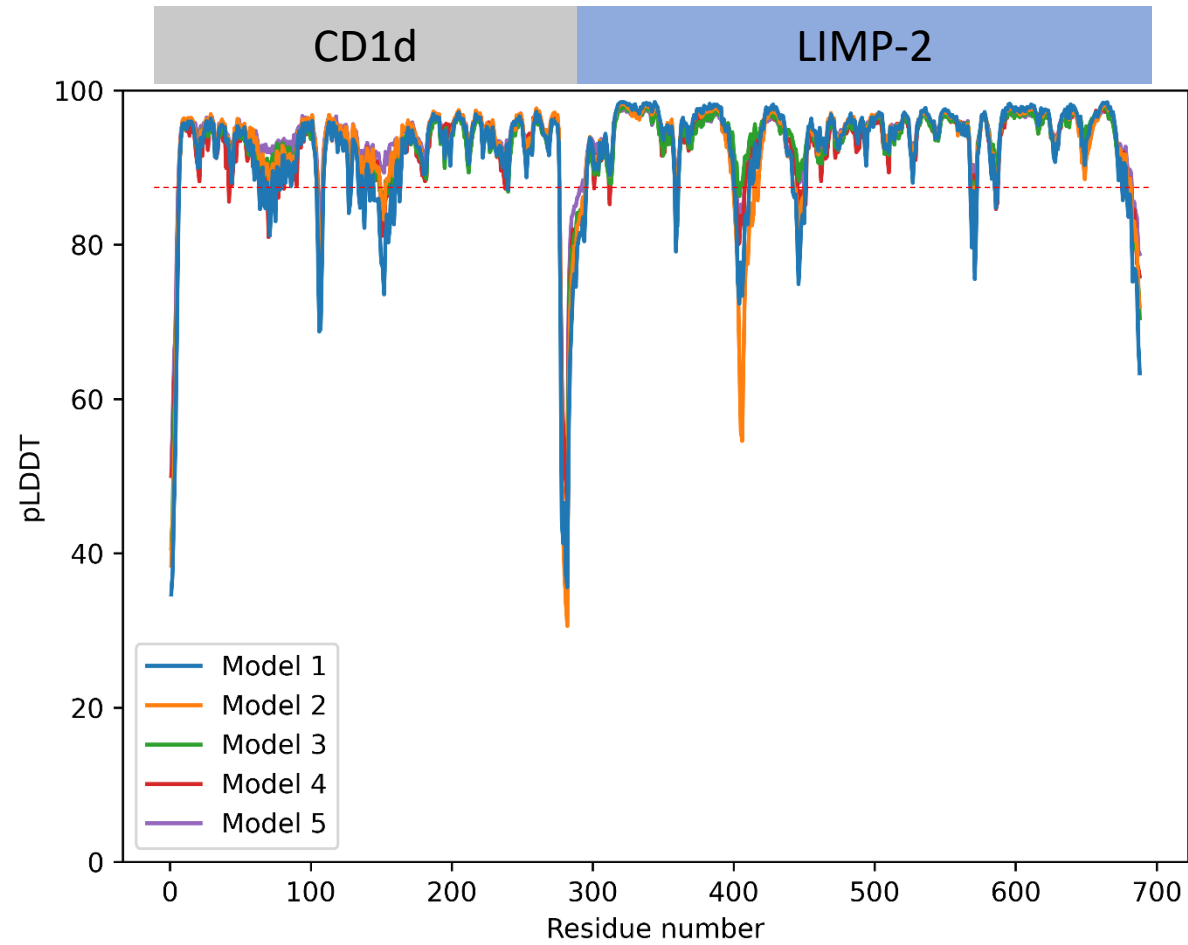
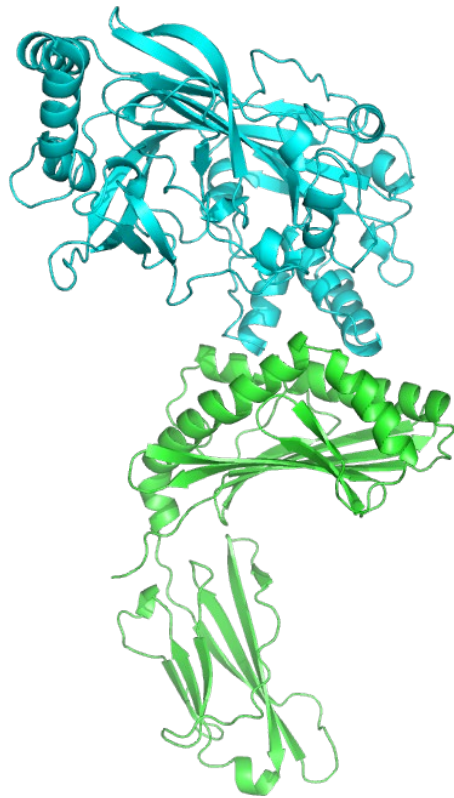
CD1d-LIMP2

- Lipid binding proteins: CD1d is like MHC-I, LIMP-2 has a lipid tunnel
- From literature they're predicted to interact
- AF2 predicts a consistent complex (all models agree)



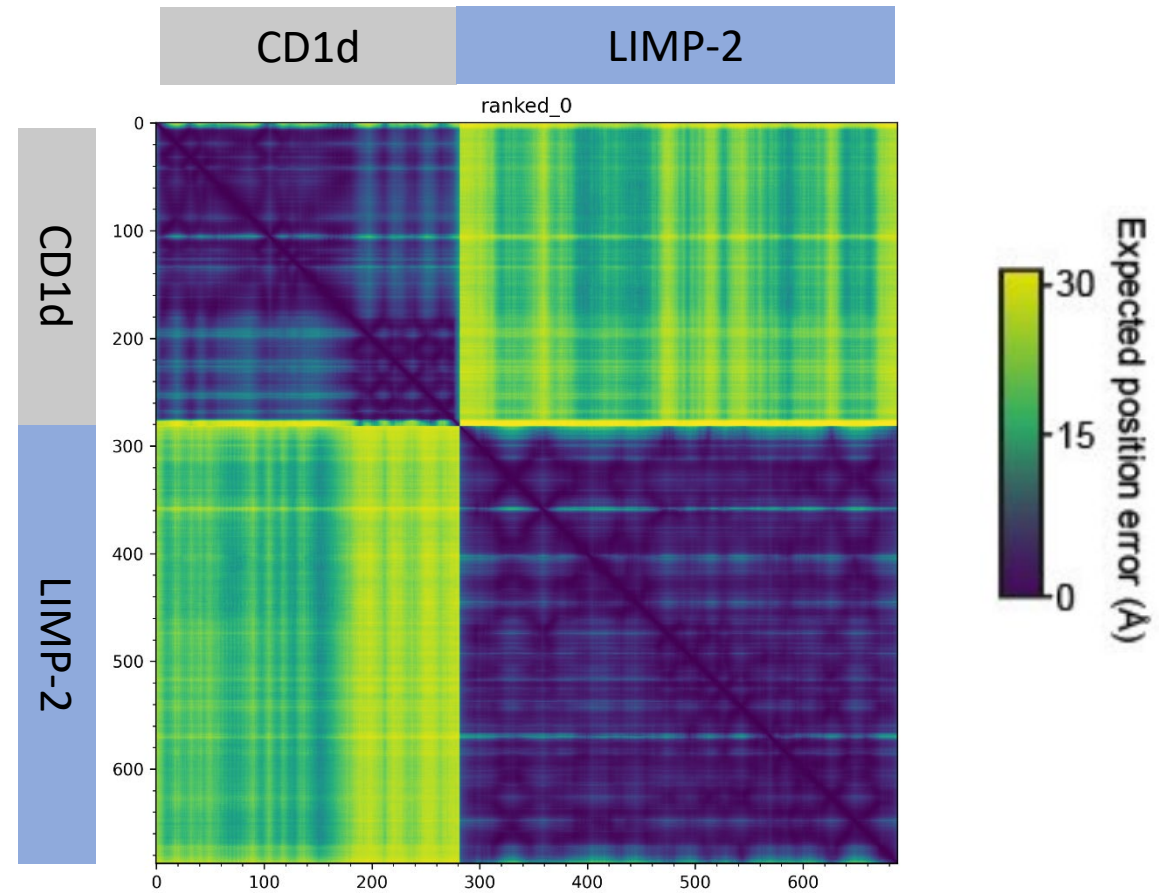
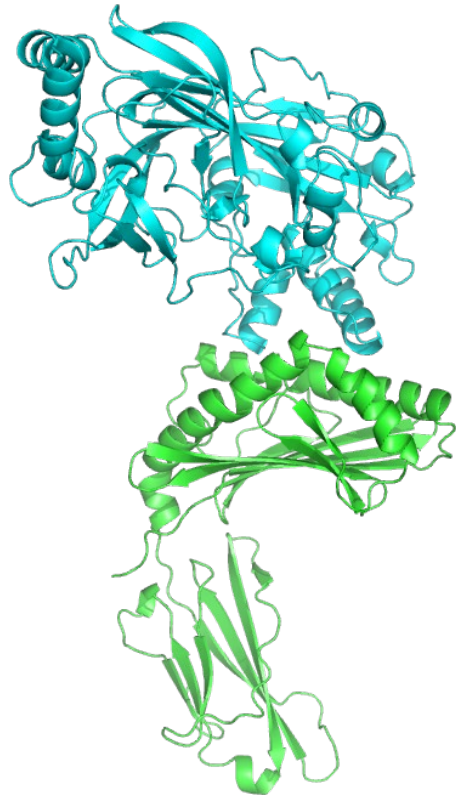
CD1d-LIMP2

- pLDDT plot



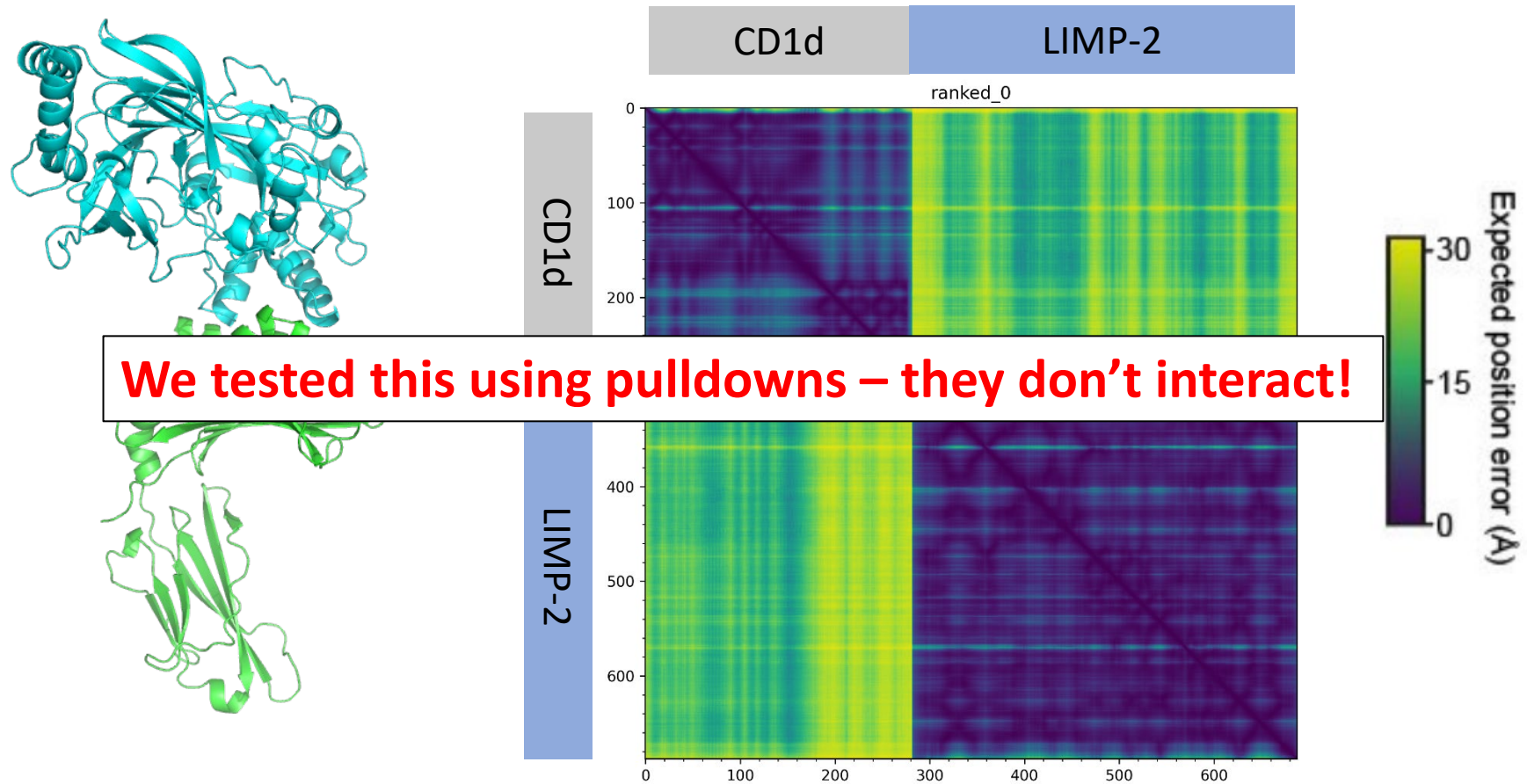
CD1d-LIMP2

- PAE plot



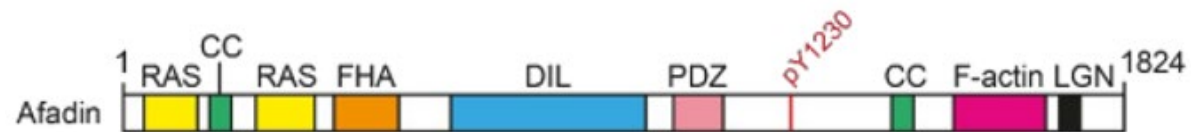
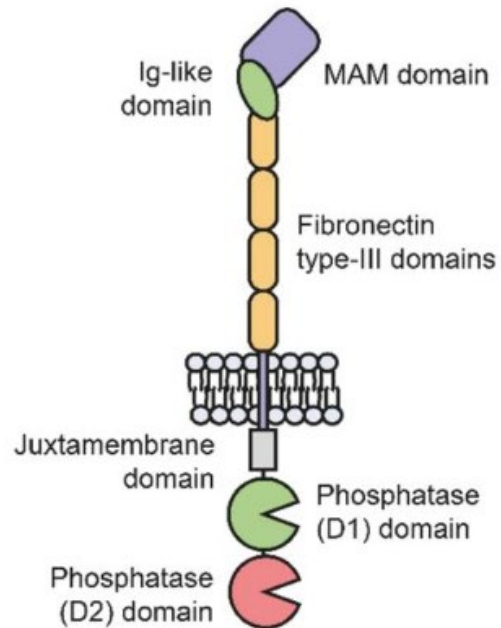
CD1d-LIMP2

- PAE plot



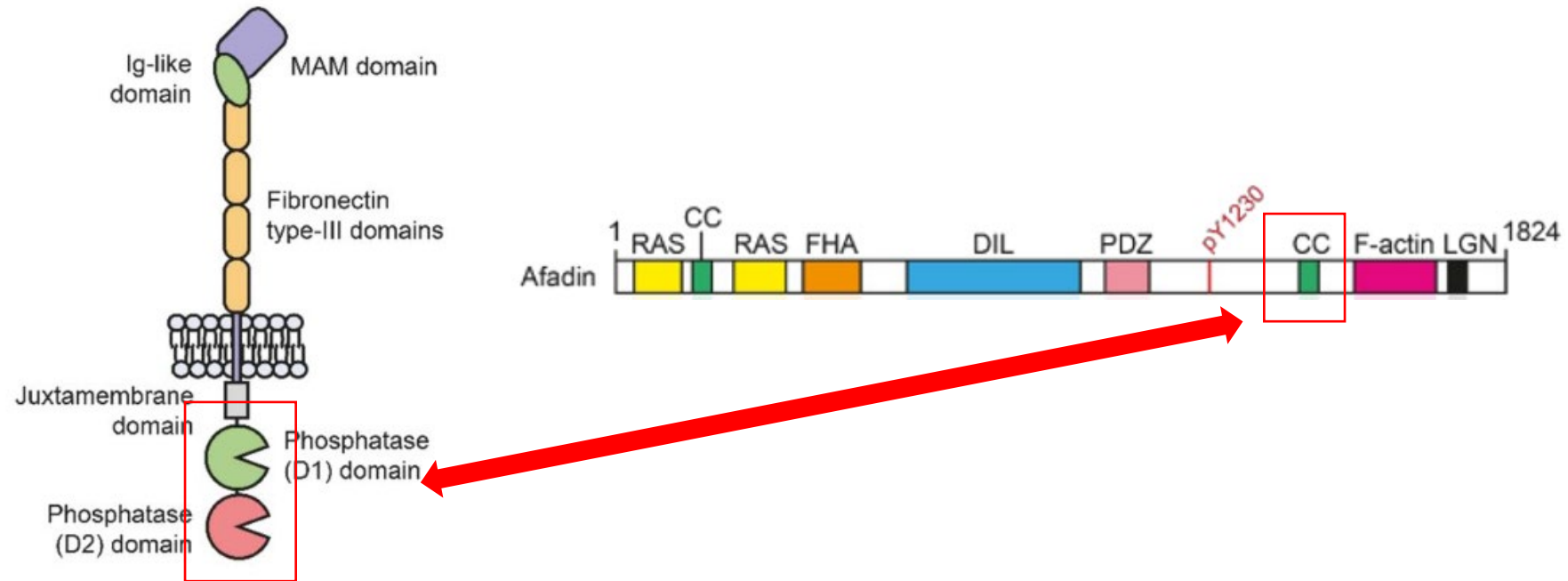
PTPRK-Afadin

- We knew that PTPRK binds Afadin but these are both BIG proteins



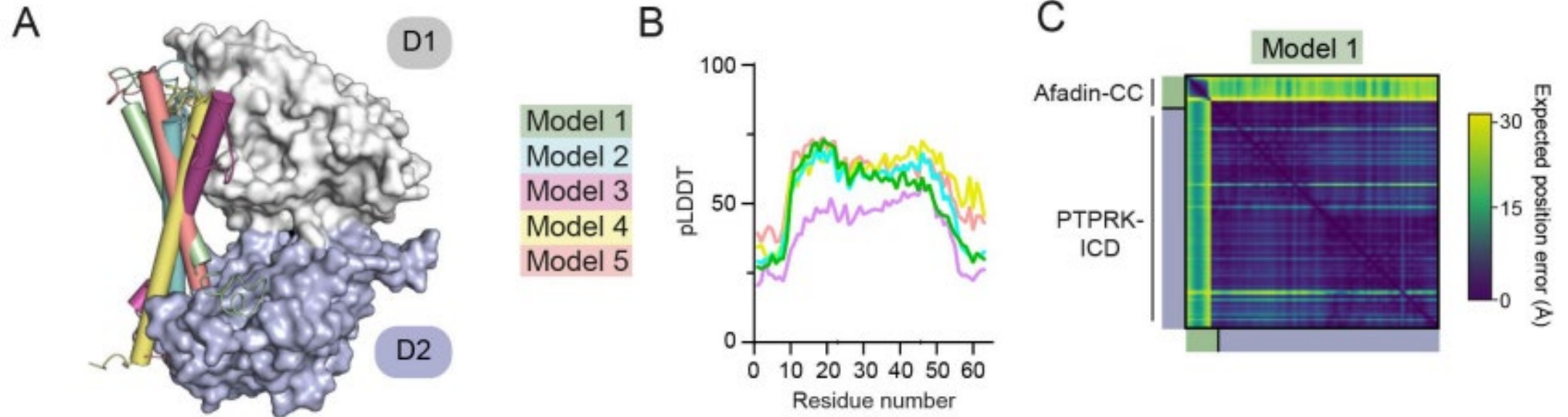
PTPRK-Afadin

- In the lab, mapped this down to much smaller domains



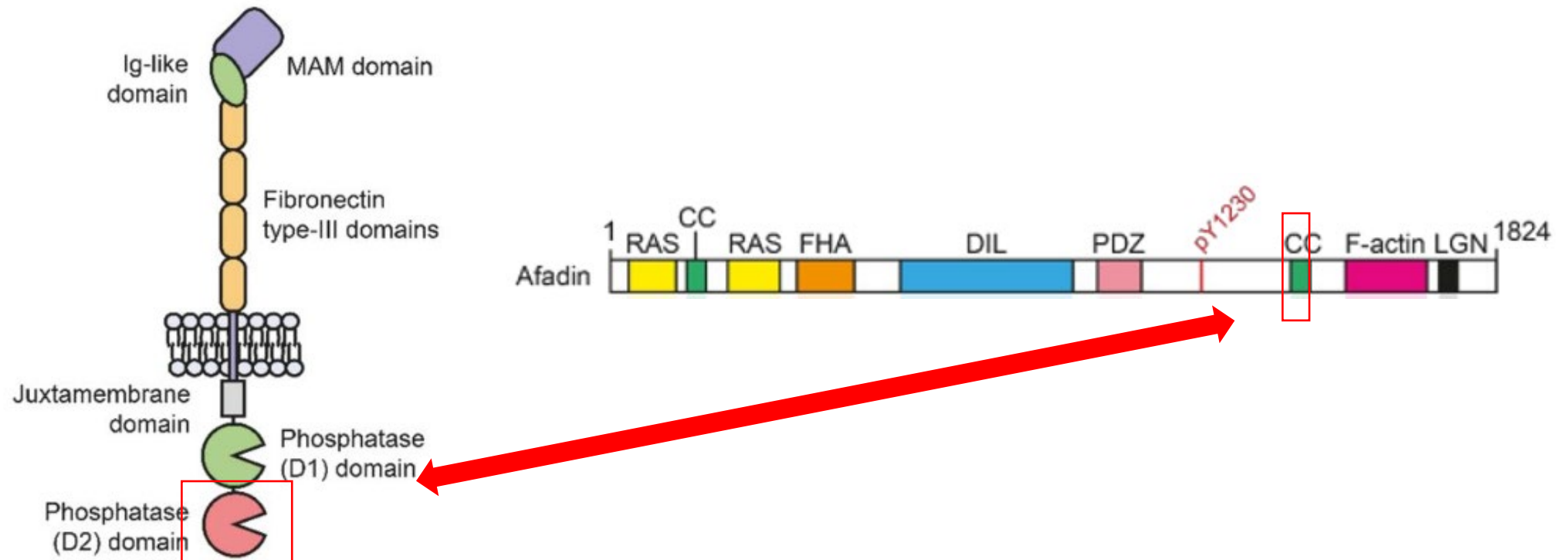
PTPRK-Afadin

- But AF2 Multimer models weren't good



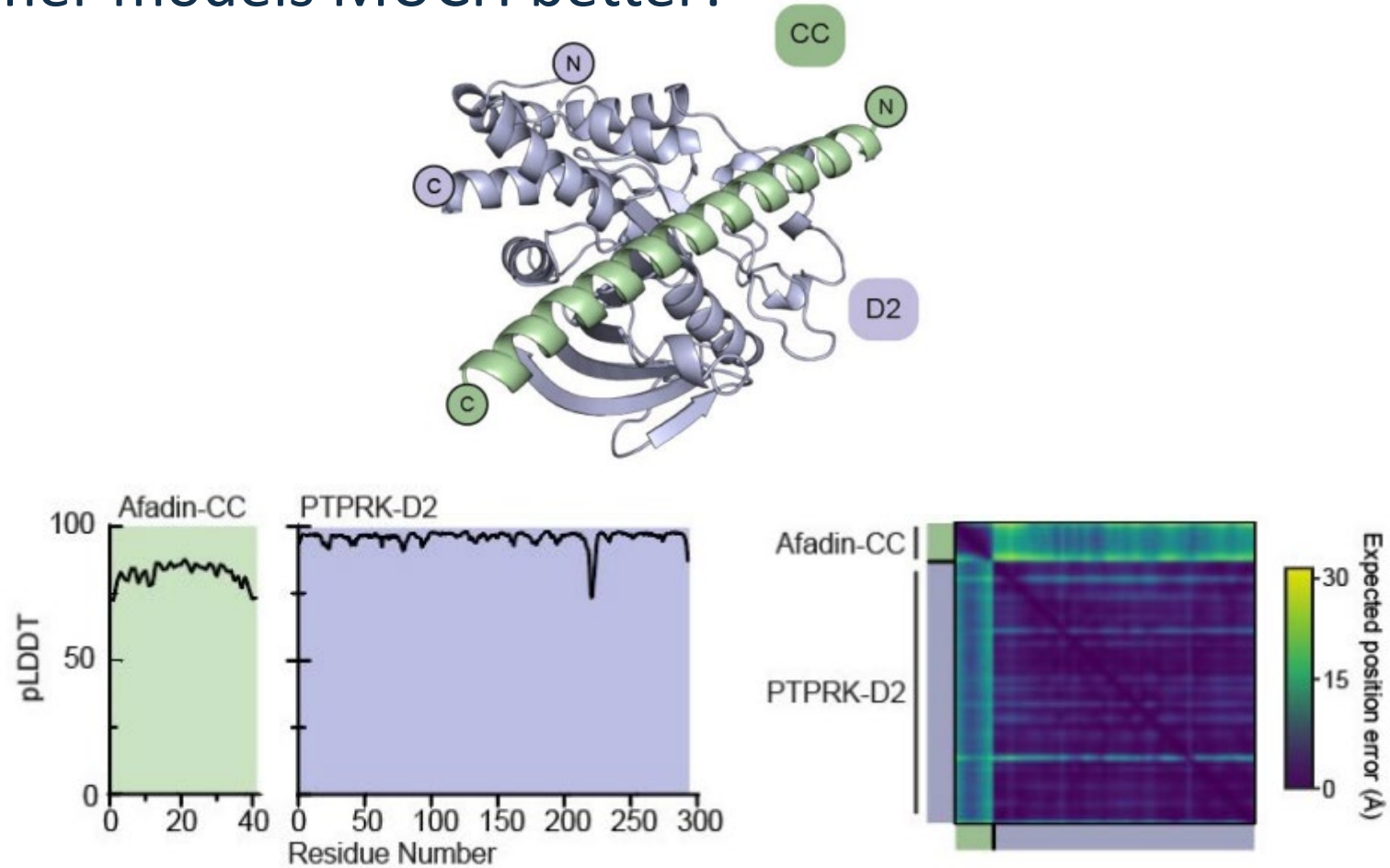
PTPRK-Afadin

- Experimentally mapped it down to smaller pieces



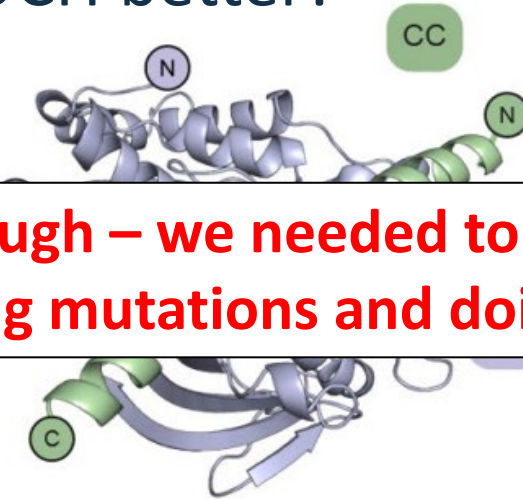
PTPRK-Afadin

- AF2 Multimer models MUCH better!

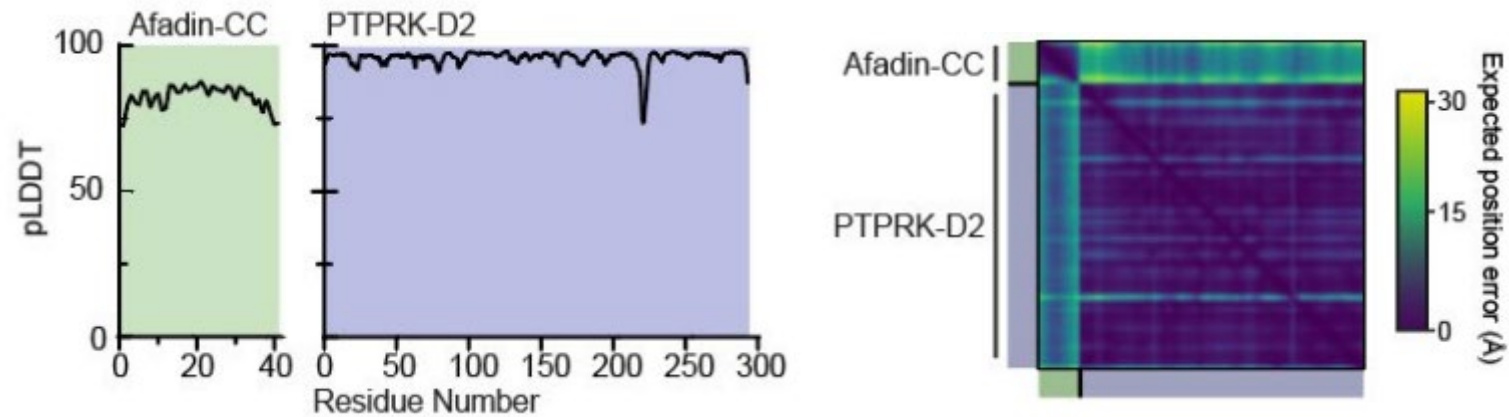


PTPRK-Afadin

- AF2 Multimer models MUCH better!

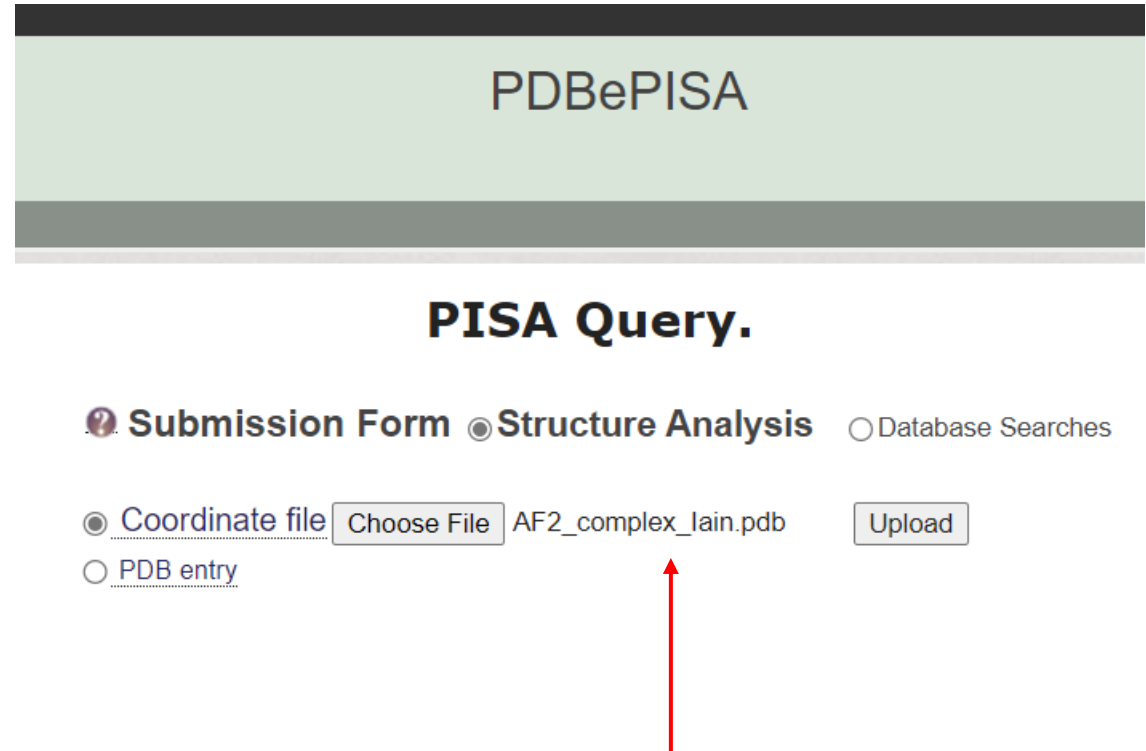


This was not enough – we needed to validate the interface by making mutations and doing pulldowns



Using PDBePISA with AlphaFold Models

- Upload AF2 model of complex to PDBePISA



PDBePISA

PISA Query.

Submission Form Structure Analysis Database Searches

Coordinate file PDB entry

Choose File AF2_complex_lain.pdb Upload



Using PDBePISA with AlphaFold Models

PISA Query.

Submission Form Structure Analysis Database Searches

[AF2_complex_lain.pdb](#) uploaded.

Coordinate file No file chosen

PDB entry

Analysis: 2 amino acid chains in ASU

Cell parameters:


A:	<input type="text" value="not given"/>	Alpha:	<input type="text" value="not given"/>
B:	<input type="text" value="not given"/>	Beta:	<input type="text" value="not given"/>
C:	<input type="text" value="not given"/>	Gamma:	<input type="text" value="not given"/>

Crystallographic information not found. You may give the cell parameters and the space symmetry group in the fields above. You may also submit without crystal data.



Using PDBePISA with AlphaFold Models

PISA Interface List.


Session Map  (id=372-60-497)




Start **Interfaces** Interface Search

Monomers


Assemblies

Interfaces in AF2_complex_lain.pdb

Interfaces  XML View Details Download Search

##	Structure 1			x	Structure 2			interface	ΔG	ΔG	N_{HB}	N_{SB}	N_{DS}	CSS
	NN 	Range	Surface \AA^2		Range	Surface \AA^2	area, \AA^2							
1	 A	81-21	4797		B	96-29	13477	829.7	1.9	0.599	8	12	0	0.000

View Details Download Search



Using PDBePISA with AlphaFold Models

Hydrogen bonds <input type="button" value="XML"/>				Salt bridges <input type="button" value="XML"/>				No disulfide bonds found
##	Structure 1	Dist. [Å]	Structure 2	##	Structure 1	Dist. [Å]	Structure 2	No covalent bonds found
1	A:ARG 23[HH11]	1.77	B:GLU 179[OE2]	1	A:ARG 23[NE]	3.84	B:GLU 179[OE2]	
2	A:ARG 23[HH21]	2.15	B:ASP 118[O]	2	A:ARG 23[NH1]	2.73	B:GLU 179[OE2]	
3	A:ARG 25[HH22]	2.01	B:GLU 223[OE1]	3	A:ARG 23[NH1]	3.29	B:GLU 179[OE1]	
4	A:ARG 25[HH21]	2.16	B:GLU 223[OE2]	4	A:ARG 23[NH2]	3.75	B:ASP 118[OD2]	
5	A:GLN 30[HE22]	1.79	B:GLU 220[OE2]	5	A:ARG 25[NH2]	2.90	B:GLU 223[OE1]	
6	A:LYS 33[HZ3]	2.13	B:GLU 221[OE1]	6	A:ARG 25[NH2]	2.81	B:GLU 223[OE2]	
7	A:GLU 22[OE1]	2.07	B:ARG 225[HH11]	7	A:LYS 26[NZ]	3.45	B:GLU 223[OE2]	
8	A:GLU 22[OE2]	1.83	B:ARG 225[HH22]	8	A:LYS 33[NZ]	2.81	B:GLU 221[OE1]	
				9	A:GLU 22[OE1]	3.07	B:ARG 225[NH1]	
				10	A:GLU 22[OE1]	3.70	B:ARG 225[NH2]	
				11	A:GLU 22[OE2]	3.56	B:ARG 225[NH1]	
				12	A:GLU 22[OE2]	2.82	B:ARG 225[NH2]	



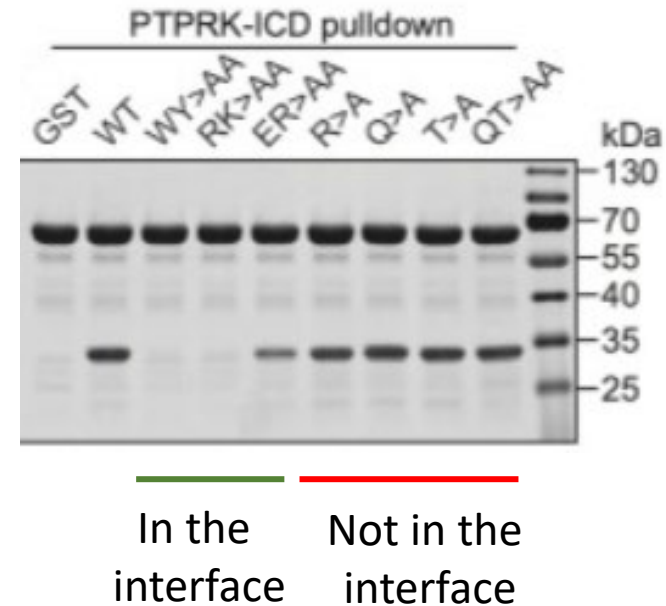
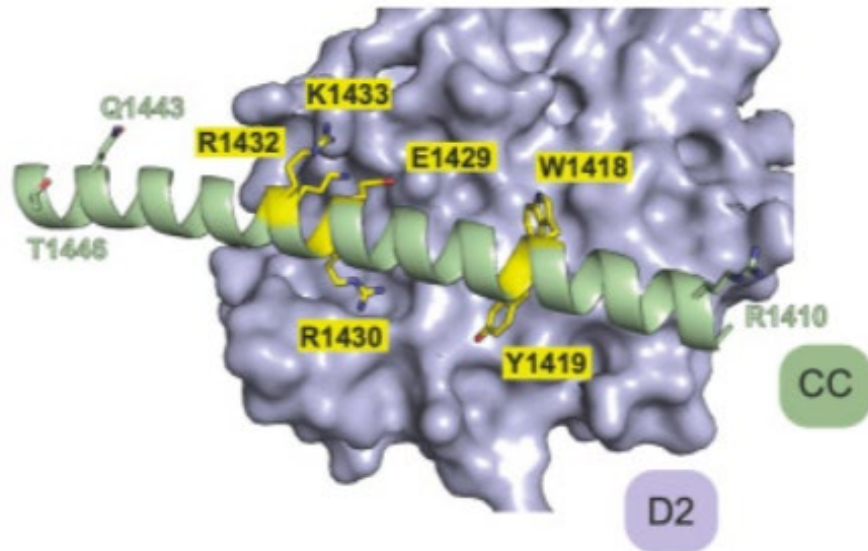
Using PDBePISA with AlphaFold Models

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PTPRK-Afadin




- Our pulldowns using mutations based on the AF2 model validated the interface experimentally



A few caveats

- PDBePISA didn't predict this interface to be significant - but it was!

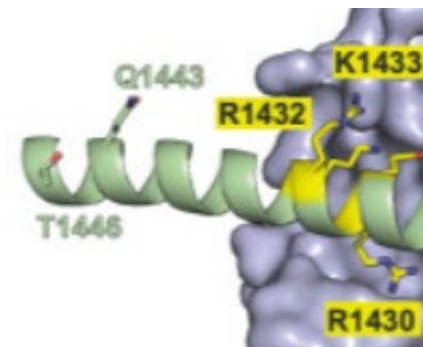
Interfaces  XML View Details Download Search

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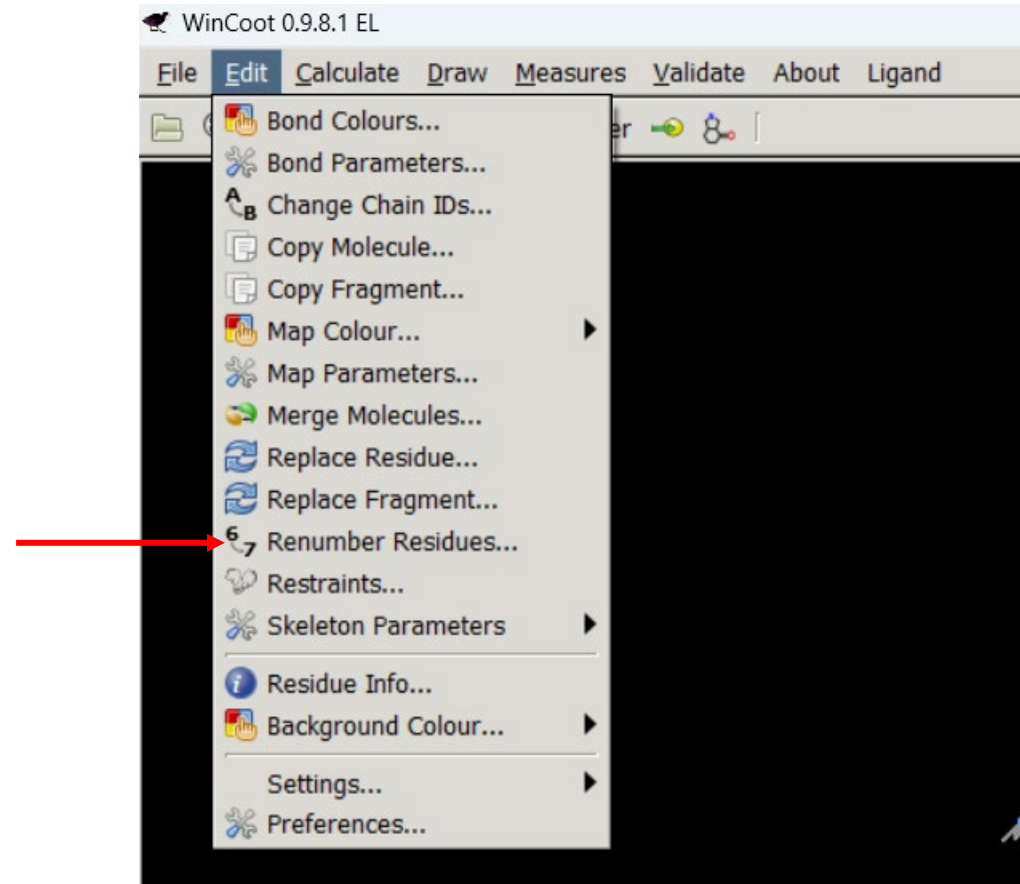
View Details Download Search

- AF2 rennumbers your residues so they might no longer match the Uniprot entry – you can renumber your model using Coot

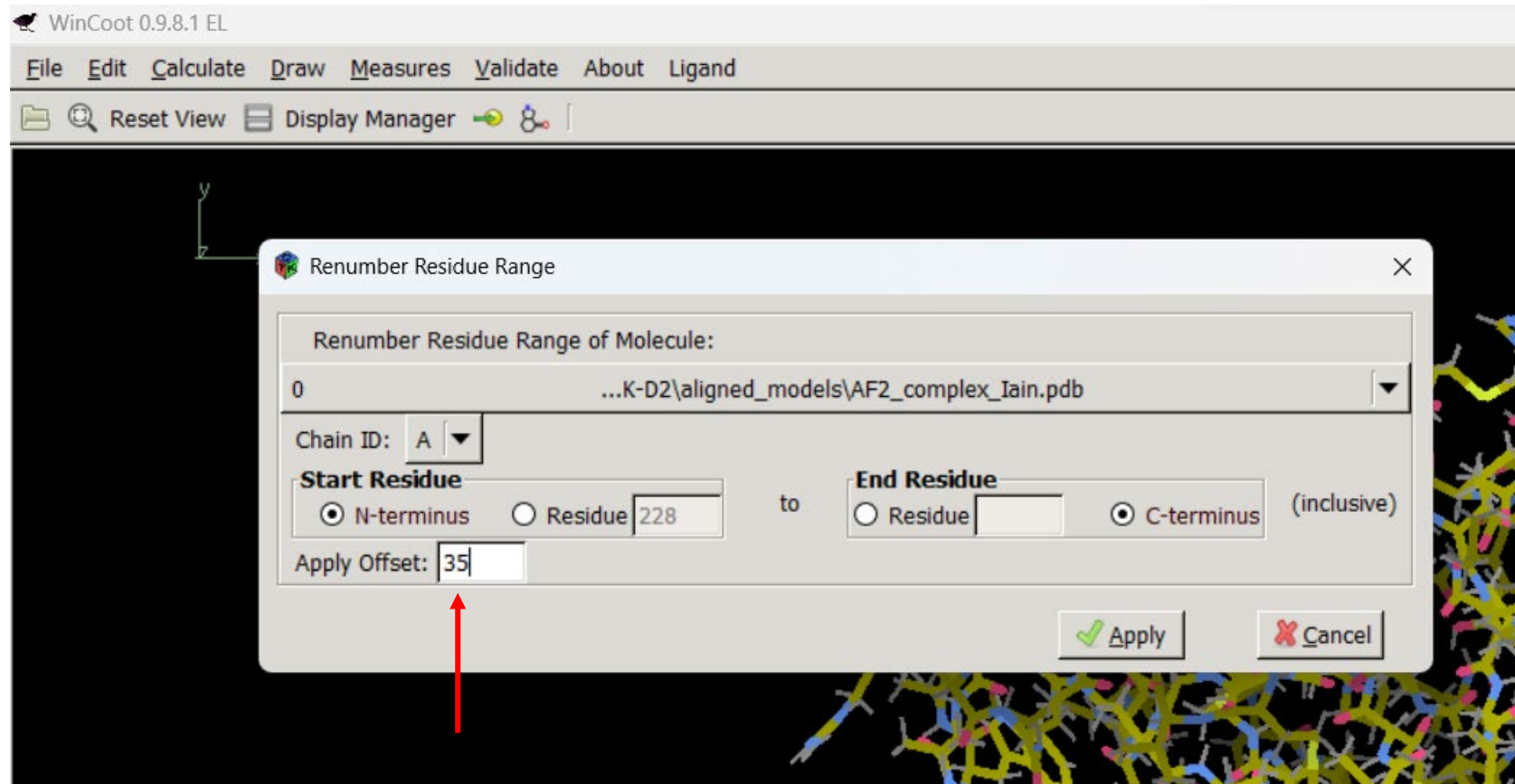
A:ARG 25[NH2] 2.81 B:GLU 223[OE2]
A:LYS 26[NZ] 3.45 B:GLU 223[OE2]



Renumber residues in Coot

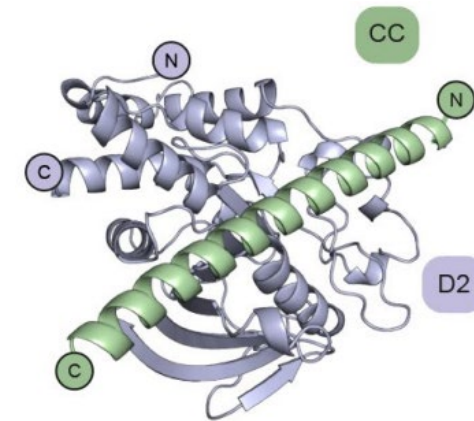
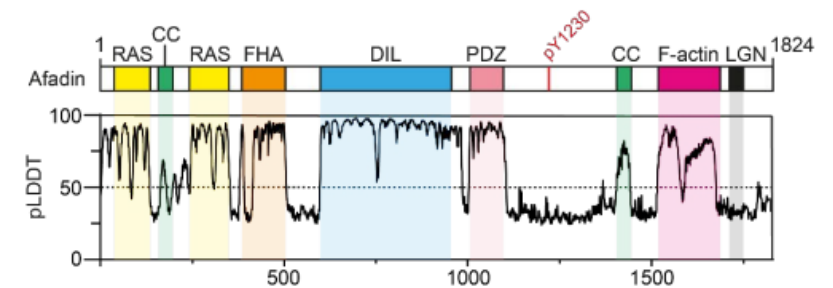
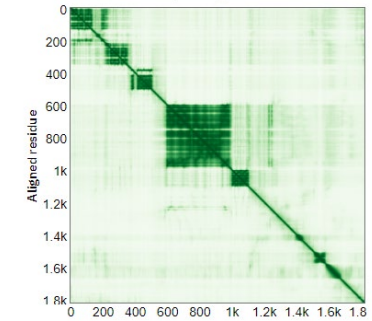


Renumber residues in Coot



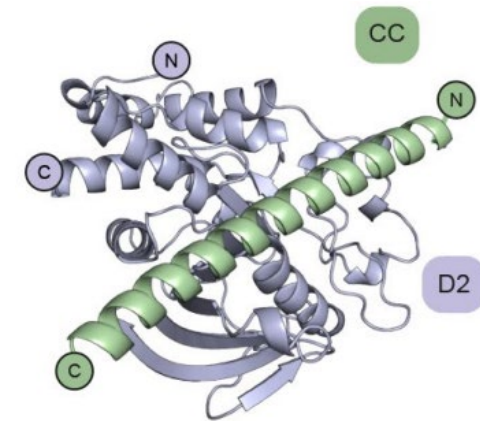
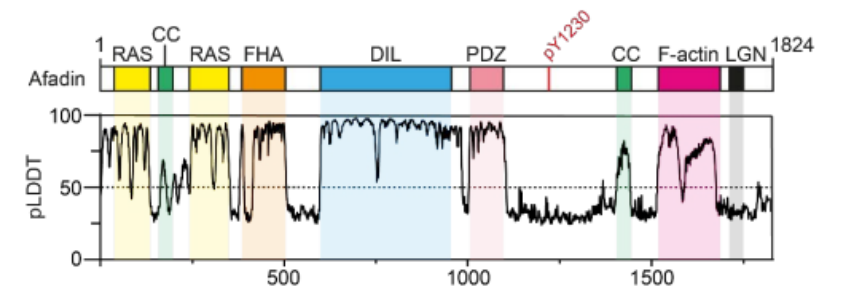
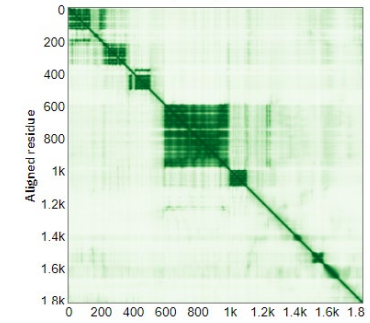
So, what is AlphaFold2 good for?

- Determining the fold of protein domain(s)
 - Identify potential functional homology
- Determining domain boundaries
 - Clone sensible constructs
- Protein:peptide complexes
 - And some protein:protein complexes



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- AF2 models should always be:
 - Shown with their statistical plot
 - Tested experimentally



What AF2 isn't good at (yet!)

- Most protein:protein complexes
 - But gives testable hypotheses
- Predicting surface properties
 - OK but not perfect, interpret with caution



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AlphaFold is being constantly developed and expanded
It is likely several of these limitations will be overcome eventually

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Try it yourself

- You can access all the pre-calculated AlphaFold structures by DeepMind/EMBL-EBI:
 - <https://alphafold.ebi.ac.uk/>
- You can run AF2 yourself via the browser (Google Colab):
 - <https://colab.research.google.com/github/deepmind/alphafold/blob/main/notebooks/AlphaFold.ipynb>



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- NOTE: if you want to run locally on your computer you need a very powerful computer (GPU with lots of RAM) and we recommend installing ColabFold not AlphaFold

